

1 Trial-by-trial inter-areal interactions 2 in visual cortex in the presence or 3 absence of visual stimulation

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14 **Abstract** State-of-the-art computational models of vision largely focus on fitting trial-averaged
15 spike counts to visual stimuli using overparameterized neural networks. However, a
16 computational model of the visual cortex should predict the dynamic responses of neurons in
17 single trials across different experimental conditions. In this study, we investigated trial-by-trial
18 inter-areal interactions in the visual cortex by predicting neuronal activity in one area based on
19 activity in another, distinguishing between stimulus-driven and non-stimulus-driven shared
20 variability. We analyzed two datasets: calcium imaging from mouse V1 layers 2/3 and 4, and
21 extracellular neurophysiological recordings from macaque V1 and V4. Our results show that
22 neuronal activity can be predicted bidirectionally between L2/3 and L4 in mice, and between V1
23 and V4 in macaque monkeys, with the latter interaction exhibiting directional asymmetry. The
24 predictability of neuronal responses varied with the type of visual stimulus, yet responses could
25 also be predicted in the absence of visual stimulation. In mice, we observed a bimodal
26 distribution of neurons, with some neurons primarily driven by visual inputs and others showing
27 predictable activity during spontaneous activity despite lacking consistent visually evoked
28 responses. Predictability also depended on intrinsic neuronal properties, receptive field overlap,
29 and the relative timing of activity across areas. Our findings highlight the presence of both
30 stimulus- and non-stimulus-related components in interactions between visual areas across
31 diverse contexts and underscore the importance of non-visual shared variability between visual
32 regions in both mice and macaques.

34 Introduction

35 To predict the activity of neurons in the visual cortex, multiple studies have focused on correlat-
36 ing external stimuli with trial-averaged responses (*Hubel and Wiesel, 1962; Pasupathy et al., 2020*).
37 Between the stimulus and specific cortical neurons, there is a complex signal processing cascade in-
38 volving multiple processing stages. Therefore, computational models of visual processing typically
39 gloss over most of the relevant biological machinery in an attempt to fit average firing rates from
40 images (*Serre et al., 2007a; Yamins et al., 2014*). A mechanistic understanding of the factors that

41 govern firing in the visual cortex requires models that can capture the trial-by-trial transformations
42 across those processing stages. Moreover, neurons throughout the cortex fire “spontaneously” in
43 the absence of any visual input. Thus, by definition, any model predicting neuronal activity that
44 is exclusively dependent on visual stimulation does not account for such fluctuations. Previous
45 studies in mice have revealed significant non-visual influences in neuronal activity in cortex, even
46 in V1, partly accounted for by movement (*Stringer et al., 2019b; Avitan and Stringer, 2022; Polack*
47 *et al., 2013; Niell and Stryker, 2010; Dadarlat and Stryker, 2017*). These observations contrast
48 with a recent macaque study which did not find the same motor-related spontaneous activity in
49 either V1, V2, or V3 (*Talluri et al., 2023*). Nevertheless, variables that are not related to movement,
50 such as attention, expectation, and arousal, also modulate stimulus- and non-stimulus driven neu-
51 ronral activity in monkeys (*Reynolds and Chelazzi, 2004; Gazzaley et al., 2007; Okazaki et al., 2008;*
52 *Gilbert and Li, 2013*), potentially adding to the response variability across stimulus repeats and to
53 neuronal activity in the absence of visual stimuli.

54 Neuronal interactions between visual areas occur in the presence and absence of visual stimuli
55 (*Chen et al., 2022; Stringer et al., 2019b; Wosniack et al., 2021; Ringach, 2009; Avitan and Stringer,*
56 *2022*). Therefore, such interactions can and should be studied both as a function of sensory inputs
57 and contextual cues but also in the absence of external stimulation or task demands (*Chacron*
58 *et al., 2003; Hsu et al., 2004; Ringach, 2009*). A paradigmatic example of inter-areal interactions
59 is the series of synaptically-connected laminar (e.g. layer 4 → layer 2/3) and cortical areas (e.g.
60 V1→V2→V4→IT) within the ventral visual stream (*Lee et al., 2016; Felleman and Van Essen, 1991;*
61 *Markov et al., 2014; Douglas and Martin, 2004; Wang and Burkhalter, 2007; Consortium et al.,*
62 *2021*). Due to feedforward, feedback, and horizontal connections in the ventral visual stream, the
63 inter-areal interactions could represent reliable shared visual and non-visual information. Several
64 studies examined interactions between visual areas in mice and macaques, focusing on the en-
65 tire population level (*Semedo et al., 2019, 2022; Tang et al., 2023; Morales-Gregorio et al., 2024*),
66 trial-averaged responses removing transient fluctuations (*Semedo et al., 2019*), neuronal activity in
67 response to only one image presentation (*Semedo et al., 2019, 2022*), or in the absence of any stimu-
68 lus (*Morales-Gregorio et al., 2024*). Here we investigated interactions between areas in single trials
69 at the level of cortical layers or brain areas across different stimulus types or in the absence of visual
70 stimulation, across different species, and across different recording techniques and temporal res-
71 olutions. Throughout, we use the term “interactions” to denote shared variability at the single-trial
72 level, including stimulus-induced correlations and trial-by-trial stimulus-independent fluctuations
73 which can arise from internal state, contextual inputs, or other top-down factors. We focused on
74 multiple simultaneously recorded areas of the ventral visual stream to assess the stimulus- and
75 non-stimulus-driven variability shared between cortical subnetworks. We found that it is possible
76 to reciprocally predict neuronal activity, both during visual stimulation but also during spontaneous
77 activity, and that this predictability depends on the intrinsic properties of each neuron, the degree
78 of receptive field overlap, and the relative timing of activity across areas.

79 Results

80 Layer 4 activity predicts layer 2/3 activity and V1 activity predicts V4 activity in 81 single trials

82 We studied neuronal activity from two open datasets: mouse neurons in V1 layer 4 and layers 2/3
83 (L4 and L2/3; calcium imaging; *Figure 1A*) (*Stringer et al., 2019a*), and macaque monkey multiunit
84 sites in areas V1 and V4 (extracellular electrophysiology; *Figure 1B*) (*Chen et al., 2022*). In addi-
85 tion, we recorded new data from an additional monkey during spontaneous activity and using the
86 same visual stimuli as in *Chen et al. (2022)* (Methods). The mouse neuronal recordings we used
87 for this experiment were based on approx. 5,500 neurons per mouse (n=4, *Table 1*) responding to
88 visual stimuli (drifting gratings or static natural black and white images; total of 7 recording days),
89 in addition to “spontaneous” activity during approximately 30 minutes of gray/black screen pre-

90 sentation on 6 of the 7 recording days. The monkey recordings were based on a range of 32-803
91 electrodes per animal (3 monkeys, L, A, and D; **Table 2**) responding to visual stimuli (full-size static
92 checkerboard image; n=3 recordings for monkey L, n=1 for monkey A, n=2 for monkey D), small
93 and thin bar slow-moving in a small clockwise square direction (n=1 for monkeys L and A); or large
94 and thick bar fast-moving in a big clockwise square direction (n=1 for monkeys L and A) in addition
95 to spontaneous activity during gray screen presentation in all recording days (n=5 for monkey L,
96 n=3 for monkey A, n=2 for monkey D). The recordings consisted of multi-unit activity (MUAe) and
97 local field potentials (LFP) in monkeys L and A retrieved from the open dataset by Chen et al. (**Chen**
98 **et al., 2022**). Monkey D recordings were acquired using the same checkerboard stimulus presen-
99 tation conditions as stated in **Chen et al. (2022)**. There was also a lights-off condition, where the
100 head-fixed monkeys were free to open or close eyes for approximately 10-25 minutes (monkeys
101 L/D; 2-3 recording days). We omitted channels with signal-to-noise ratio of less than 2, or channels
102 that were considered “spurious” by the authors of the open dataset (**Chen et al., 2022**).

103 We evaluated two types of interactions between areas: inter-laminar (**Figure 1C**; mouse V1) and
104 inter-cortical (**Figure 1D**; monkey). We used linear ridge regression to predict neuronal activity in
105 one area from activity in the other area in single trials (**Figure 1E**, see **Methods**) (**Semedo et al., 2019**).
106 For each neuron/site in the target area we *fit* a linear readout from the other area’s simultaneous
107 population activity on the same trial, with an L_2 (“ridge”) penalty. Performance was evaluated using
108 cross-validation over trials and quantified as squared Pearson’s r (hereafter, “explained variance”
109 or EV, **Methods**). **Figure 2A** shows sample neuronal activity from three example mouse V1 L2/3 cells
110 during image presentation (black traces). Overlaid, the figure also shows the predicted neuronal ac-
111 tivity (red). The predicted neuronal activity is shown as a function of the actual activity in response
112 to every image presentation for the same example cells in **Figure 2C**. The top cell illustrates a case
113 where the predicted activity closely matches the actual activity ($EV = 0.67$), the middle cell shows a
114 typical case ($EV = 0.39$), and the bottom cell illustrates a case where the predictions deviated from
115 the actual neuronal activity ($EV = 0.07$). We focused on neurons deemed “visually reliable” (~17%
116 of total L2/3 neurons; **Table 1**, **Methods**, see results for all neurons in **Figure Supplement 1C**). The
117 ridge regression model predicted single-trial L2/3 activity from L4 activity across both types of vi-
118 sual stimuli with an average EV of 0.28 ± 0.16 (mean \pm stdev. across neurons, **Figure 2E**) whereas
119 the shuffle control mean EV was 0.004 ± 0.002 .

120 In monkeys, trial-to-trial variations in V4 activity were predicted from V1 activity across the three
121 types of visual stimuli. Example recording sites for monkey L are shown in **Figure 2B, D** (see also **Fig-**
122 **ure Supplement 1 D–G** for examples from two additional monkeys, A and D). The ridge regression
123 model predicted the single-trial responses in V4 activity from V1 activity with an average EV of 0.30
124 ± 0.15 (**Figure 2F** for monkey L, see **Figure Supplement 1H** for monkey A and **Figure Supplement 1I**
125 for monkey D), whereas the shuffle control mean EV was 0.005 ± 0.005 . Most sites in V4 (72.3%)
126 were deemed visually reliable; EV results for all monkey sites are shown in **Figure Supplement 1J**).
127 EV performance varied across the three monkeys ($EV_L = 0.34$, $EV_A = 0.21$, $EV_D = 0.11$). This varia-
128 tion may be due to predictor population size (**Table 2**). We subsampled sites in monkey L to match
129 the numbers of monkey A and monkey D (20 permutations; **Figure Supplement 1L**; subsampling
130 was performed for all analyses hereafter unless noted). Subsampled monkey L matching the num-
131 ber of sites in monkey A (monkey L_A) showed an average EV of 0.35 ± 0.14 , statistically higher
132 compared to the EV for monkey A ($p < 0.001$, permutation test). Similarly, subsampled monkey
133 L matching the number of sites in monkey D (monkey L_D) showed an average EV of 0.19 ± 0.11 ,
134 statistically higher compared to the EV for monkey D ($p < 0.001$, permutation test).

135 We asked whether the conclusions were dependent on the model and parameter choices. As an
136 alternative approach to ridge regression, we also fit a Poisson general linear model (GLM) enforc-
137 ing non-negativity to monkey MUAe without baseline subtraction, using identical folds, bin widths,
138 and temporal gaps as in the ridge regression analysis. The results from the Poisson GLM were
139 consistent with the results obtained using ridge regression (**Figure Supplement 1M**). Additionally,
140 to evaluate the impact of the bin size on the results, we re-estimated EV after binning MUAe into

141 10–200 ms. EV increased with bin size and remained well above shuffle controls across conditions
142 (**Figure Supplement 4A–C**, $p < 0.001$, paired permutation test), indicating that inter-areal predictabil-
143 ity is robust across a broad range of physiologically relevant timescales.

144 In sum, it was possible to provide estimates of neuronal activity in single trials in both species,
145 across different layers within primary visual cortex in mice and across different visual cortical areas
146 in monkeys.

147 **Inter-cortical predictions are asymmetrical**

148 In the previous section, we demonstrated the possibility of predicting L2/3 activity from L4 activ-
149 ity and V4 from V1. We asked whether we could also predict neuronal responses in the opposite
150 direction. We could predict L4 activity from L2/3 activity in mice and V1 activity from V4 activity in
151 monkeys (**Figure Supplement 1A** for mice, **Figure Supplement 1K** for monkeys). To directly compare
152 predictability between directions in mice and monkeys, we matched the number of predictors (i.e.,
153 number of neurons/sites used to predict activity) and the degree of self-consistency (split-half r val-
154 ues) by randomly subsampling in each layer or cortical region prior to computing the predictability
155 metrics (**Figure 3A, C, Methods**).

156 In mice, there was no statistically significant difference between L4→L2/3 and L2/3→L4 direc-
157 tions ($p > 0.05$, hierarchical permutation test, **Figure 3B**). In monkeys, after matching predictor
158 count and split-half correlation values, the EV fraction in the V1→V4 direction was higher than in
159 the V4→V1 direction ($p < 0.001$ for monkey L, **Figure 3D**; $p < 0.05$ for monkey A, **Figure Supple-**
160 **ment 3C**). This directional asymmetry was stable across temporal bin sizes in monkey L (**Figure**
161 **Supplement 4D**). In monkey A, the asymmetry was small and was not robust across different bin
162 sizes (**Figure Supplement 4E**), consistent with the noisier data and weaker effect size in that ani-
163 mal. There were not enough sites with similar reliability distributions in monkey D to perform the
164 matched subsampling directionality tests.

165 Although we accounted for predictor size and reliability distributions, differences in predic-
166 tion performance between directions could still reflect additional inherent differences in target-
167 population properties (**Figure Supplement 2F–J**). To control for these differences, we modeled EV
168 as a function of target-population properties and used the residuals for inference, $EV_{\text{resid}} = EV - \widehat{EV}$.
169 Covariates included self-consistency, SNR, one-vs-rest r^2 , and variance metrics (**Methods**). After ad-
170 justment, the difference between inter-cortical directions remained statistically significant overall
171 ($p < 0.001$ for monkey L, $p < 0.001$ for monkey A, **Figure Supplement 3F–G**). These results suggest
172 that the differences between directions cannot be attributed to target-related neuronal proper-
173 ties. We also analyzed the residuals in mice prediction directions, and after controlling for neuron
174 properties, we still found no statistically significant difference in predictability directions (**Figure**
175 **Supplement 3E**, $p = 0.22$ for stimulus activity, $p = 0.48$ for gray screen activity).

176 **Predictability of neuronal activity is dependent on the visual stimulus**

177 We evaluated whether the predictability of neuronal activity varied with the type of visual stimulus
178 presented to the animal. In mice, we compared the inter-laminar prediction of neuronal activity
179 of visually reliable neurons in response to drifting gratings versus natural images (**Figure 4A**). We
180 could predict mouse L4 and L2/3 activity under both stimulus conditions ($p < 0.001$, paired permuta-
181 tion test of prediction vs. shuffled frames prediction). Predictability was higher for drifting gratings
182 than natural images in the L4→L2/3 direction (**Figure 4B**; $p < 0.001$, hierarchical permutation test).

183 In monkeys L and A, we compared inter-cortical predictability of visually reliable sites in re-
184 sponses to a slow-moving small thin bar, fast-moving large thick bar, and a full-size checkerboard
185 image (**Figure 4C**). As expected, V1 and V4 could predict each other's neural activity across all stim-
186 ulus types ($p < 0.001$, paired permutation test of prediction vs. shuffled frames prediction). The
187 predictability was highest in both directions for neuronal activity in response to a full-field checker-
188 board image (monkey L **Figure 4D**; monkey A **Figure Supplement 5F**). In the V1→V4 direction, the

189 EV fraction was higher when predicting a slow-moving small thin bar compared to a fast-moving
190 large thick bar (monkey L *Figure 4D*, left; monkey A *Figure Supplement 5F*, left).

191 In the V4→V1 direction, there was no consistency in prediction differences between monkey L
192 and monkey A. In monkey L (V4→V1), the EV for fast-moving large thick bars was larger than for
193 the slow-moving thin bars ($p < 0.001$, permutation test; *Figure 4D*, right), whereas this difference
194 was flipped in monkey A ($p < 0.001$; *Figure Supplement 5F*, right). This observation could be due to
195 differences in the degree of reliability in the neuronal responses between monkeys (*Figure Supple-*
196 *ment 5H*, L for monkeys L, A, respectively). We modeled EV as a function of SNR, self-consistency
197 (split-half r), variance across time within stimulus, and variance across trials within timepoint, and
198 used $EV_{\text{resid}} = EV - \widehat{EV}$ for inference (Methods). In monkey L (V4→V1), the difference in EV resid-
199 uals between thin and thick bars was not statistically significant ($p = 0.44$; *Figure Supplement 5K*,
200 right), whereas this difference remained statistically in monkey A ($p < 0.001$; *Figure Supplement 5O*,
201 right). Lastly, in the V1→V4 direction, EV residuals remained statistically higher when predicting
202 the responses to the thin bar compared to the thick bar in both monkeys L and A ($p < 0.001$ for
203 both monkeys; *Figure Supplement 5K,O*, left).

204 **Neuronal activity could be predicted even during spontaneous activity**

205 Given the dependence on the visual stimulus, we next asked whether it would be possible to predict
206 neuronal responses in the absence of any visual stimulus, during “spontaneous activity”. We com-
207 pared the predictability of stimulus-evoked activity in mice (drifting gratings and natural images)
208 versus the predictability of activity recorded during gray screen presentations. This comparison
209 was conducted in both visually (SNR >2 & split-half $r > 0.8$) and non-visually (SNR <2 & split-half
210 $r < 0.8$) reliable neurons (n=3 mice; mouse MP027 did not undergo 30 min. of gray screen pre-
211 sentation). In visually reliable neurons, neuronal activity could still be predicted during the gray
212 screen condition ($p < 0.001$, hierarchical paired permutation test compared with shuffled frames).
213 There was a significant reduction in EV during gray screen compared to visual stimulus presenta-
214 tion (*Figure 5A* left, $p < 0.001$, hierarchical paired permutation test). In non-visually reliable neurons,
215 predictability was higher during the gray screen condition compared to stimulus presentation (*Fig-*
216 *ure 5A* right, $p < 0.001$, hierarchical paired permutation test).

217 While there was no correlation between neuronal predictability in the responses to visual stim-
218 ulus presentations and in the response to gray screen presentations in visually reliable neurons
219 (*Figure 5B*), there was a strong correlation for non-visually reliable neurons (*Figure 5C*). The differ-
220 ence in predictability in the absence of a stimulus (gray screen presentations) could in principle
221 change according to the directionality in inter-laminar interactions. However, there was no statisti-
222 cally significant difference in the EV fraction between laminar directions (L4→L2/3 vs. L2/3→L4), in
223 the same “visually reliable” group and even when subsampling “non-visual” neurons (*Figure Sup-*
224 *plement 3A*).

225 In monkeys, we only focused on visually reliable sites since the majority of the neuronal popu-
226 lation was visually reliable (*Figure Supplement 1J*). Additionally, an SNR of less than 2 (one of the
227 requirements to define non-visual neurons in the mouse data) most likely reflects artifactual is-
228 sues with the electrode recording the multiunit site (*Chen et al., 2022*). We compared inter-areal
229 prediction of stimulus presentation activity (all monkeys for checkerboard presentation; monkeys
230 L and A for moving bars), gray screen presentation (all monkeys), and during lights-off (monkeys
231 L and D). The predictability of neuronal activity in response to gray screen presentation remained
232 statistically above chance ($p < 0.001$ paired permutation test of prediction vs. shuffled frames pre-
233 diction for all monkeys). The predictability of neuronal activity in the absence of visual stimulus
234 was lower compared to checkerboard presentations (*Figure 5D*, $p < 0.001$ for monkey L; *Figure*
235 *Supplement 6E*, $p < 0.001$ for monkey A; *Figure Supplement 6F* left, $p < 0.01$ for monkey D). The
236 same conclusions held when comparing moving bar vs. gray screen presentations (*Figure Sup-*
237 *plement 6D* for monkey L; *Figure Supplement 6E* for monkey A). Intriguingly, the EV fraction in
238 the lights-off condition was statistically higher than during the stimulus presentations in both di-

239 rections in monkey L, whereas the EV fraction in the lights-off condition in monkey D was lower
240 (*Figure Supplement 6F*). Eye closure and sleep can induce global oscillations (*Hohaia et al., 2022*)
241 and therefore may lead to correlated neuronal activity, causing an increase in predictability. Mon-
242 key L spent a significant portion with its eyes closed (*Chen et al., 2022*), unlike monkey D, who spent
243 almost the entire time with its eyes open (eyes closed duration < 3s in both recording days). To
244 test the effects of eye closure, we further separated the lights-off neuronal activity of into periods
245 where the eyes of monkey L were open or closed. The EV fraction was statistically significantly
246 higher than EV fraction during stimulus presentation activity only during the eyes-closed period
247 (*Figure 5D*).

248 The correlation in visual predictability between stimulus presentation and spontaneous activity
249 was high across all types of spontaneous conditions (*Figure 5E* and *Figure Supplement 6C* for mon-
250 key L; *Figure Supplement 6I, J* for monkeys A, D, respectively). When assessing the inter-cortical
251 prediction directionality during spontaneous conditions, in monkey L, we found the same asym-
252 metrical relationship as in *Figure 3*, where V1→V4 EV fraction was significantly higher than V4→V1
253 prediction in both gray screen ($p < 0.001$, permutation test) and lights-off ($p < 0.001$, permutation
254 test) conditions (*Figure Supplement 3B*). In monkey A gray screen presentation activity, we did not
255 find any statistically significant difference between direction predictions (*Figure Supplement 3C*,
256 right). However, when accounting for neuronal property differences between the areas, we found
257 the same direction EV asymmetry across both monkeys ($p < 0.001$; *Figure Supplement 3F* for mon-
258 key L; *Figure Supplement 3G* for monkey A).

259 **Behavioral metrics account for only a small portion of predictability**

260 Because inter-areal predictability persists in the absence of visual drive, we asked whether be-
261 havioral signals could partly account for the inter-laminar and inter-cortical interactions. In mice,
262 augmenting the neural predictability model with face-motion singular value decomposition (SVD)
263 components and running speed did not significantly increase EV relative to the neural-only model
264 in the L2/3 to L4 direction ($p = 0.77$, paired permutation test), and had a mild statistical increase
265 in the L4 to L2/3 direction ($p < 0.05$, paired permutation test; *Figure Supplement 10A*). During
266 spontaneous activity, the same type of augmentation did not increase EV relative to the neural-
267 only model in either direction ($p = 0.095$ for L4→L2/3 direction, $p = 0.93$ for L2/3→L4 direction,
268 paired permutation tests; *Figure Supplement 10C*). Behavior-only models captured a modest yet
269 statistically significant fraction of variance (stimulus: $EV=0.02 \pm 0.03$, spontaneous: $EV=0.01 \pm 0.01$;
270 $p < 0.001$ paired permutation test with shuffled frames). Behavior-only EV correlated with neu-
271 ral EV within areas (stimulus: $\sim 0.47-0.50$; spontaneous: $\sim 0.68-0.69$; *Figure Supplement 10B,D*),
272 indicating that animals/neurons with stronger neural predictability also tended to show stronger
273 behavior-related structure, but in all cases behavior was not driving the main effect.

274 In monkey neural recordings during resting state condition (eyes open only), pupil diameter
275 provided a small but significant predictive power ($EV=0.01 \pm 0.01$; $p < 0.001$ paired permutation
276 test of prediction vs. shuffled frames prediction). Adding pupil size to the neural model yielded no
277 significant EV gain in either direction ($p = 1$ in the V1→V4 direction, $p = 0.70$ in the V4→V1 direction;
278 *Figure Supplement 10E*).

279 Together, these results indicate that behavioral metrics contributed a measurable shared com-
280 ponent to cross-area predictability, but they are not the principal driver: most explained variance
281 remains tied to the neural population activity.

282 **Receptive field overlap and neuronal response properties impact predictability**

283 We investigated which neuronal properties are related to the ability to predict responses by com-
284 paring EV and key indicators of neuronal response reliability and receptive field properties, in both
285 visually- and non-visually reliable neurons, during either visual presentation or spontaneous condi-
286 tions. First, we considered the following properties: (i) max r^2 value (i.e., maximum squared corre-
287 lation between each neuron in the predictor population and the predicted neuron), (ii) 1-vs-rest r^2

288 self-consistency (squared correlation between each neuron's response to one trial repetition of all
289 stimuli/timepoint with the average response of the rest of the trial repetitions; *Methods*), (iii) SNR
290 of the predicted neuron, (iv) variance across stimuli, (V) variance across repeats of the same stimu-
291 lus, (vii) variance across timepoints (monkey data only, since we have multiple timepoints per trial),
292 and (viii) split-half r (*Methods*). We plotted EV against each of these variables (mouse: **Figure 6B**;
293 monkey L: **Figure 6E**, monkey A: **Figure Supplement 8B**, monkey D: **Figure Supplement 9B**) and
294 report the correlation coefficient between EV and each variable in the y-axis in mice (**Figure 6A**)
295 and monkeys (monkey L: **Figure 6D**, monkey A: **Figure Supplement 8A**, monkey D: **Figure Supple-**
296 **ment 9A**).

297 In mice, during both stimulus presentation and gray screen presentation, the most correlated
298 property with a neuron's inter-areal predictability was the max r^2 (**Figure 6A**). For the other 5 prop-
299 erties, there was a strong distinction between stimulus presentation (dark bars) and gray screen
300 presentation (light bars): all 5 properties were positively correlated with the neural activity pre-
301 dictability EV fraction during stimulus presentation, but they were slightly anticorrelated with their
302 predictability EV fraction during gray screen presentation. Because the split-half correlation calcu-
303 lation averages out the non-stimulus-dependent variability in both halves of the trials, it showed
304 a weaker correlation with EV, which depends on trial-by-trial modulation. The one-vs-rest r^2 met-
305 ric, which also examines trial-by-trial modulation and does not average split-half trials, yielded a
306 stronger correlation with EV during stimulus presentation.

307 When examining the relationship between 1-vs-rest self-consistency and inter-laminar predic-
308 tion EV in mice, we observed a bimodal distribution of neurons: one group of neurons showed
309 high EV despite having low self-consistency, and another group showed EV correlated with self-
310 consistency (**Figure 6B** third column; **Figure Supplement 7A**; $p < 0.001$, Hartigan's dip test after
311 thresholding neurons with $EV > 0.4$). The responses of neurons with low self-consistency also
312 showed high EV during gray screen presentation. This bimodality was present in two of the three
313 mice ($p < 0.001$ for mouse MP031, MP032, $p = 0.69$ for mouse MP033, Hartigan's dip test; self-
314 consistency and EV fraction relationships across all mice can be seen in **Figure Supplement 7B**). To
315 better understand the responses of neurons with low self-consistency, we projected out the "non-
316 visual ongoing neuronal activity" from the neuronal responses (**Stringer et al., 2019a**) (*Methods*).
317 This non-visual ongoing activity is deemed to be influenced by spontaneous behavior (**Stringer**
318 **et al., 2019b**). Projecting out this non-visual activity eliminated this bimodality ($p = 0.71$ and $p = 0.98$
319 for L4→L2/3 and L2/3→L4, respectively; Hartigan's dip test, **Figure 6C,D**). This observation could be
320 because the responses of neurons with low self-consistency can no longer be predicted, or be-
321 cause the responses of those neurons became more reliable and therefore were highly predicted.
322 To distinguish between these two possibilities, we compared both the 1-vs-rest self-consistency
323 and the prediction EV before and after removing the non-visual activity. Removing the non-visual
324 ongoing activity increased the self-consistency value across the three mice (**Figure Supplement 7C**;
325 $p < 0.001$, paired permutation test). Interestingly, the inter-laminar EV fraction decreased in MP031
326 and MP032 mice, yet increased in MP033 (**Figure Supplement 7F** $p < 0.001$, paired permutation test).
327 When examining individual pairwise relationships in a fraction of highly predictable neurons (lines
328 in **Figure Supplement 7F**), we found that some of the highly predictable neurons remained pre-
329 dictable after removing the non-visual activity, whereas other highly predictable neurons dropped
330 their EV fraction dramatically. Together with the small yet reliable shared behavior contributions
331 (**Figure Supplement 10**), these results indicate that a non-trivial portion of inter-areal predictability
332 in mice reflects shared internal-state fluctuations, whereas a complementary portion persists after
333 removing those dimensions, consistent with stimulus-driven interactions.

334 In monkeys, one of the neuronal properties that showed high correlation with inter-areal pre-
335 diction EV across all conditions was also the maximum correlation value (first column in **Figure 6D**,
336 **E**, **Figure Supplement 8A**, and **Figure Supplement 9**). Other neuron properties like SNR, split-half
337 correlation, and one-vs-rest correlation were also highly correlated with inter-cortical predictabil-
338 ity EV (**Figure 6D**, **Figure Supplement 8A**, and **Figure Supplement 9A**). The split-half correlation was

339 highly correlated with EV fraction, although the relationship was highly non-linear (middle column
340 in *Figure 6D*, *Figure Supplement 8B*, and *Figure Supplement 9B*). Using the one-vs-rest squared
341 correlation removed some of this non-linearity and further increased the correlation with the EV
342 fraction in monkeys L and A (third column in *Figure 6D*, *Figure Supplement 8B*). In addition, the
343 relationship between one-vs-rest correlation squared and EV fraction did not show evidence of
344 bimodality ($p = 0.99$, Hartigan's dip test focusing on sites with $EV > 0.4$).

345 We conjectured that neurons that have overlapping receptive fields (RFs) should share more
346 information, and therefore their responses would better predict each other than neurons with
347 non-overlapping RFs. In addition, even when all neurons are exposed to the same stimulus (full
348 field symmetrical checkerboard image, gray screen, darkness, etc), neurons with overlapping RFs
349 may be more synaptically connected, resulting in better inter-cortical predictions. To test this hy-
350 pothesis, we compared inter-cortical predictions in different ensembles of neurons with RFs that
351 differed in their degree of overlap. This hypothesis was only tested in monkeys L and A because
352 we did not have access to RF estimates in the mouse data, and we did not have enough sites with
353 RF overlap in monkey D. For each V4 site whose responses we predicted, we separated the pre-
354 dictors into two groups with matched size: one group where all the V1 predictor sites had <10%
355 RF overlap (sample of one V4 site in monkey L, *Figure 6F*, top), and one group where all the V1
356 predictor sites had >80% RF overlap (sample of one monkey L V4 site, *Figure 6F*, bottom). A similar
357 procedure was followed when predicting the activity of V1 neurons from V4 predictor neurons (*Fig-
358 ure 6H*). Inter-areal prediction was higher in the >80% RF overlap condition compared to the <10%
359 RF overlap ensembles in both directions and across all stimulus conditions (*Figure 6G*, I for monkey
360 L; *Figure Supplement 8C*, D for monkey A). To compare these numbers with those obtained with
361 larger numbers of sites, we calculated the EV performance while using all predictors. In most cases,
362 predictions using the >80% RF overlap group were lower than when using all predictors, showing
363 that predictor neurons with low RF overlap still contributed to the EV (*Figure 6G*, I, for monkey L;
364 *Figure Supplement 8C*, D for monkey A).

365 **Inter-areal predictability is both stimulus and non-stimulus driven**

366 The results in *Figure 5* and *Figure 6* pointed to components of the predictable responses that are
367 stimulus-driven and other components that are not stimulus-driven. We considered two alterna-
368 tive hypotheses: (1) If shared information were strictly stimulus-driven, prediction across repeats
369 of the same stimulus should remain accurate; (2) If shared information were entirely stimulus-
370 independent, prediction across repeats should approach chance ($EV \approx 0$). To test these hypothe-
371 ses, we compared the inter-areal prediction EV fractions using unshuffled versus shuffled trials.
372 Shuffling was performed across repeat trials of the same images (mice: *Figure 7A*, monkeys: *Fig-
373 ure 7D*). In mice, one stimulus presentation was either a drifting grating or a natural static image.
374 In monkeys, one stimulus presentation was either the one checkerboard image, a large, thick, fast-
375 moving bar, or a small, thin, slow-moving bar. In both species and in both directions, inter-areal
376 prediction EV fraction decreased after shuffling stimulus repeats compared to before shuffling
377 (*Figure 7B* for mouse; *Figure 7E* for monkey L; *Figure Supplement 11A* for monkey A; *Figure Sup-
378 plement 11C* for monkey D; $p < 0.001$ for both species and directions, paired permutation test).

379 Interestingly, the EV fraction during shuffled trial-repeats still remained significantly above
380 chance ($p < 0.001$, paired permutation test of prediction vs. shuffled frames prediction). In mice,
381 neurons showed two types of distribution in terms of their response predictability in shuffled and
382 unshuffled trials. For a subset of neurons, the EV fraction was still high in the shuffled condition,
383 albeit their EV was lower than in the unshuffled case (*Figure 7C*; points below but near the diago-
384 nal line). For another subset of neurons, the EV fraction during shuffled trials was much lower or
385 even near zero. The responses of the latter group had the highest predictability during gray screen
386 activity.

387 In monkeys, neurons farther away from the diagonal line showed higher EV fraction during
388 gray screen activity (*Figure 7F* for monkey L; *Figure Supplement 11F* for monkey A; *Figure Supple-*

389 **ment 11G** for monkey D). In addition, we examined whether shuffling repeat presentations of gray
390 screen images (simulating spontaneous activity) would result in any prediction at all. There was a
391 more profound decrease in inter-cortical performance in all monkeys (**Figure Supplement 12A–C**)
392 with no neurons that remained as predictable during shuffled trials compared to unshuffled trials
393 (**Figure Supplement 12E–G**).

394 **Accounting for latency differences improves inter-areal activity predictions in mon-** 395 **keys visual area sub-populations**

396 Given the latency differences in neuronal responses between V1 and V4 *Schmolesky et al., 1998*,
397 we asked whether accounting for this latency could result in better inter-cortical prediction. To test
398 this hypothesis, we offset the neuronal activity using different lags for each area (**Figure 7G, H**) and
399 recalculated the ridge regression predictions. For each offset level, we calculated the percentage of
400 neurons where the EV fraction peaked at that offset. For the checkerboard image, in monkeys L and
401 A V1→V4 predictions, the biggest percentage of neurons had a peak performance when there was
402 no time offset between areas, with a substantial proportion of neurons with a peak performance
403 for 10–30 ms offsets in the negative direction (i.e., V1 activity preceding V4 activity; **Figure 7I**, left for
404 monkey L, **Figure Supplement 11E**, left for monkey A). This distribution of peak EV values was only
405 present during early visual responses (first 275 ms of stimulus onset). In monkeys L and A V4→V1
406 direction, there was a large proportion of neurons with peak EV when considering 10–20 ms offsets
407 in the positive direction (i.e., V4 after V1, **Figure 7I**, right for monkey L, **Figure Supplement 11E**, right
408 for monkey A). These differences were apparent in the early part of the visual response, before 250
409 ms. When offsetting the neuronal responses to gray screen presentations, across all times and
410 areas, the highest percentage of neurons with peak EV was when there was no time offset (**Figure**
411 **Supplement 12D**).

412 **Neural signals can also be predicted at the ensemble level using local field poten-** 413 **tial signals**

414 We repeated the analyses in monkeys L and A using the local field potential (LFP) signals. We eval-
415 uated whether the predictability of LFP signals across areas depended on the signal frequency by
416 considering band-limited LFP amplitudes (Low: 2–12 Hz; Beta: 12–30 Hz; Gamma: 30–45 Hz; High-
417 gamma: 55–95 Hz; Hilbert envelopes, z-scored). In both monkeys, the Gamma band showed a
418 feed-forward signature in the early evoked period: the V1→V4 predictability peaked at negative
419 offsets (~10–30 ms; V1 leads), and the V4→V1 predictability peaked at positive offsets of similar
420 magnitude (**Figure Supplement 13A,D** for monkey L; **Figure Supplement 13B,E** for monkey A), con-
421 sistent with previous findings (*Semedo et al., 2022*; *van Kerkoerle et al., 2014*; *Bastos et al., 2015*).
422 The Low and Beta frequency bands exhibited a broader structure with less consistent peaks in
423 predictability offset timing.

424 **Discussion**

425 Neuronal activity in one brain region or layer within the visual cortex can be used to predict neu-
426 ral activity in another nearby and anatomically connected region or layer in single trials (**Fig-**
427 **ure 2**). In monkeys, predictability was asymmetric: V1 activity better accounted for V4 activity than
428 vice versa (**Figure 3, Figure 7**). This inter-areal prediction persisted across different stimuli (**Figure 4**)
429 but also in the absence of a visual stimulus, during gray-screen and lights-off periods (**Figure 5**). The
430 degree of predictability increased with signal-to-noise ratio, response variance, and the degree of
431 overlap between receptive fields (**Figure 6**).

432 In line with other studies in mice (*Stringer et al., 2019b*; *Niell and Stryker, 2008*; *Andermann*
433 *et al., 2011*), we observed an approximately bimodal distribution of neuronal responses, with a
434 large subset of neurons that do not show reliable responses to visual stimuli both in L4 and L2/3
435 (**Figure 6B**, third column; **Figure Supplement 7A**). However, even for the subset of neurons con-
436 sidered to be “non-visual”, at least within the set of stimuli and conditions examined here, their

437 activity remains predictable. This bimodal distribution disappears when projecting out potential
438 non-sensory ongoing activity (*Figure 6C; Figure Supplement 7D*) (*Stringer et al., 2019b, 2021*). At
439 the population level, neuronal encoding subspaces in mouse visual cortex have been shown to
440 have little overlap between visual sensory and non-sensory (behavioral) information, with only
441 one shared dimension (*Stringer et al., 2019b*). The visually unreliable, yet highly predictable, sub-
442 set of neurons described here could be the neuronal group driving this orthogonality. As expected,
443 the activity of “visual” neurons can be better predicted during visual presentation and not during
444 gray screen presentation. In stark contrast, the activity of non-visual neurons can be predicted
445 even better during gray screen presentation than during visual stimulation. There was no such
446 bimodal distribution in the data from monkeys. One possibility is that there may be no (or very
447 few) non-visual neurons in monkey V1 or V4. Indeed, the overwhelming majority of recording sites
448 in V1 and V4 responded strongly to visual stimulation. Yet, the comparisons between the results in
449 mice and monkeys reported here need to be interpreted with caution because the two datasets dif-
450 fer in terms of recording techniques (electrophysiology versus two-photon imaging), consequently
451 also in their temporal resolution (one millisecond versus hundreds of milliseconds), and the type
452 of interaction studied (across areas versus across layers), in addition to any differences between
453 species.

454 In monkeys, sites where the receptive fields (RF) of V1 and V4 overlap can better predict
455 each other compared to other sites showing little RF overlap. This observation could reflect RF-
456 dependent intrinsic connectivity between areas, but also RF-dependent shared inputs from other
457 areas, such as the thalamus. In the latter case, those putative shared inputs cannot be strictly
458 dependent on visual inputs, given that the effect of RF overlap persists even during gray screen
459 conditions.

460 Many computational models that aim to explain neuronal activity in the visual cortex are based
461 on feedforward signal propagation, with increased receptive field sizes, selectivity, and feature
462 invariance along the visual hierarchy (*Serre et al., 2007b; Kreiman, 2021; Connor et al., 2007*). Con-
463 sistent with this idea, we described an asymmetry in the degree of predictability, with V1 neurons
464 explaining V4 responses better than the other way around. This observation persisted after control-
465 ling for neuronal count and split-half correlation values and was also apparent during the lights-off
466 condition. In contrast, there was no asymmetry when comparing inter-laminar prediction direc-
467 tions in mice. The lack of asymmetry in inter-laminar prediction directions in mice could be due
468 to the slow dynamics in calcium imaging, the lack of a clear inter-areal hierarchy, or differences
469 between species.

470 The asymmetry in directionality was also observed when implementing temporal delays to inter-
471 areal prediction, consistent with processing delays across areas (*Semedo et al., 2022; Gokcen et al.,*
472 *2022; Schmolesky et al., 1998*). A substantial proportion of neurons increased their inter-areal pre-
473 dictability when offsetting the times between areas, specifically in the direction that aligns their
474 neuronal activities. In contrast to the temporal delays associated with processing visual stimula-
475 tion, during gray screen presentation, the activity of most neurons was best predicted in the ab-
476 sence of any time offsets, suggesting that the internally generated neuronal activity during spon-
477 taneous conditions may be largely driven in a non-feedforward manner. Further evidence sup-
478 porting the distinction between visually-driven and non-visually-driven interactions comes from
479 the observation that trial repeat shuffling reduced, but did not eliminate, predictability in both
480 mice and monkeys. In mice, when comparing shuffled versus unshuffled activity, we encountered
481 again a bimodal distribution, where a group of neurons was closer to the diagonal line (their re-
482 sponses were predicted as well during the shuffled compared to the non-shuffled condition), and
483 another group of neurons that were closer to the x-axis (their responses could not be predicted
484 during the shuffled condition). The responses of the latter group were best predicted during gray
485 screen activity, suggesting that they mostly shared non-visual information. The predictive power
486 in mouse V1 from layer 4 to layer 2/3 during spontaneous conditions has been recently shown in
487 *Papadopouli et al. (2024)*, consistent with our findings. The overall area population decrease in

488 predictability after shuffling may be due to the influence of non-visual activity, such as movement
489 (*Stringer et al., 2019b*), especially since these non-visual stimulus effects have been shown to occur
490 on a timescale of approximately one second, consistent with the results in our study. In macaques,
491 context-dependent effects are likely not due to movement, since the monkeys maintained fixation
492 during the stimulus task, and visually-evoked activity was not driven by movement (*Talluri et al.,*
493 *2023*).

494 The neural recordings in mice and monkeys are different in terms of: (i) recording modalities
495 (calcium versus electrophysiology), (ii) temporal resolution (hundreds of milliseconds versus mil-
496 liseconds), (iii) neuronal types, (iv) SNR, (v) cortical targets (layers versus areas), (vi) sample sizes,
497 (vii) stimuli, and (viii) task conditions (*Stringer, 2018; Chen et al., 2022*). The goal of this study was
498 not to make any direct quantitative comparison across species, but rather to investigate the prop-
499 erties of inter-areal interactions within each species.

500 Several limitations in this study are worth considering in future work. First, neither calcium imag-
501 ing nor MUAe recordings capture the timing of single spikes in individual neurons. One of many
502 ways in which the timing of individual spikes could be important is that synchronous or close-to-
503 synchronous inputs can have a larger impact on post-synaptic responses (*Salinas and Sejnowski,*
504 *2001*). Second, this study focused on linear predictability, but other non-linear models could better
505 capture neuronal activity. Third, and critically, the experimental data evaluated here provide only
506 a fraction of the inputs to a given neuron. For example, the predictor signals studied here are likely
507 to exclude most (in monkeys) if not all (in mice) inhibitory inputs (*Jiang et al., 2015; Shen et al., 2020;*
508 *Ibrahim et al., 2020; Schuman et al., 2021*). Any cortical neuron receives extensive local inputs and
509 non-local inputs from many other areas. Fourth, we only had limited access to non-visual sources
510 that could contribute to predicting neuronal responses; other sources of input could include finer
511 behavioral measurements, attention, familiarity, and arousal state. Finally, biophysically realistic
512 models of the transformation between inputs and outputs of a given neuron should include their
513 dendritic locations and specific synaptic potentials (*Park et al., 2019; Petousakis et al., 2023*). There-
514 fore, the quantitative results on the prediction of neuronal responses reported here constitute a
515 lower bound.

516 We evaluated inter-areal interactions in different types of neuronal recordings, timescales, and
517 species. These interactions were assessed in single trials, separating visually-driven and non-visual
518 contributions, and accounting for the directionality and dynamics of neuronal responses. These
519 efforts constitute an initial step towards, and a lower bound for, systematically building compu-
520 tational models that can account for the transformations from sensory inputs to their encoding
521 across multiple processing steps in the cortex. Whereas many efforts with artificial neural net-
522 works merely map inputs to outputs (*Kreiman, 2021*), a detailed and systematic understanding of
523 individual processing steps can lead to better and biologically more relevant computational models
524 of sensory processing.

525 **Methods**

526 **Datasets.** We used the mouse dataset from (*Stringer et al., 2019a*) containing calcium-imaging
527 activity measurements from 43,630 neurons in layer 4 (L4) and 12,060 neurons in layers 2/3 (L2/3)
528 in V1 of 4 mice during 32 different randomly interleaved presentations of either drifting gratings
529 or gray-scale natural images (each one repeated more than 90 times), along with spontaneous ac-
530 tivity during 30 minutes of exposure to a gray/black screen (*Figure 1A*, data acquisition details in
531 *Stringer et al. (2019a,b)*). Calcium imaging activity was recorded during stimulus presentations at
532 a scan rate of 2.5 Hz or 3 Hz (each frame was acquired every 400 ms or 333 ms). The computed
533 stimulus responses per stimulus presentation were averaged based on two frames immediately
534 post stimulus onset. Cortical layers were determined using the 10-12 planar z-positions retrieved
535 during the multi-plane calcium activity acquisition. For stimulus-response and spontaneous record-
536 ings, neuronal activity of each neuron was z-scored using its 30-minute gray screen spontaneous

537 activity (mean gray-screen activity subtracted and divided by gray-screen activity standard deviation).
538

539 We used the dataset from *Chen et al. (2022)* for monkeys A and L and conducted new neural
540 recordings from one additional monkey (monkey D). The Chen et al. dataset consists of envelope
541 multiunit activity (MUAe) and LFP from 1,024 recording sites in one monkey (monkey L) in response
542 to either multiple-day recordings of more than 60 repetitions of a full-size checkerboard image,
543 moving small-thin or large-thick bars in 4 directions, gray screen presentations, or more than 30
544 minutes of baseline activity where the monkey was in a room with the lights off (*Figure 1B*). We
545 also used data from a second monkey (monkey A) from the same published dataset. However,
546 due to the degraded state of the electrodes at the time of the recording, we were only able to
547 obtain enough data from V4 sites for responses to the full-size checkerboard image, moving bars,
548 and gray screen presentations. Neuronal activity was summed over 25 ms non-overlapping bins.
549 Activity duration was 300 ms, 400 ms, and 1 s for gray screen, checkerboard, and moving bar
550 presentations, respectively. For the recordings during visual stimulation, the neuronal activity was
551 normalized by subtracting the mean activity during the gray screen presentations separately for
552 each site.

553 We collected new data from an additional monkey in response to the full-size checkerboard
554 image presentations, gray screen presentations, and lights-off activity (~15 minutes). Monkey D
555 (12 years old, 14.4 kg) was chronically implanted with floating multielectrode arrays (Microprobes
556 for Life Sciences, MD) targeting areas including V1 and V4. Arrays consisted of 16 channels per area,
557 yielding a total of 32 electrodes (V1–16, V4–16). All procedures received ethical approval by Harvard
558 Medical School Institutional Animal Care and Use Committees and conformed to NIH guidelines
559 provided in the Guide for the Care and Use of Laboratory Animals. All relevant ethical regulations
560 for animal and nonhuman primate testing and research were followed.

561 The same conditions of stimulus presentation (image/square size, presentation time, etc) were
562 presented as in *Chen et al. (2022)*. Stimulus presentation and data acquisition (including electro-
563 physiology and eye tracking) were coordinated using MonkeyLogic2 software (Hwang et al., 2019)
564 and OmniPlex Neural Recording Data Acquisition Systems (Plexon Inc.), integrated via custom MAT-
565 LAB scripts. Online spike sorting was performed with the PlexControl client using waveform-based
566 classification. Visual stimuli were displayed on ViewPixx EEG monitors (ViewPixx Technologies;
567 1920×1080, 120 Hz), and eye position was monitored via ISCAN (ISCAN Inc.). Raw analog signals
568 were used to calculate MUAe using same processing logic as *Chen et al. (2022)*.

569 **Visual Reliability.** A neuron or site was defined to be visually reliable if its signal-to-noise ratio
570 (SNR) was 2 or higher and its split-half correlation value was 0.8 or higher. Due to the high number
571 of repetitions of visual stimuli, the split-half correlation was skewed toward high values, which is
572 why we used a higher split-half correlation threshold than commonly used in other studies. In mice,
573 the SNR for each neuron was calculated as:

$$SNR_{mouse} = \frac{\langle r_{stim} \rangle - \langle r_{spont} \rangle}{std(r_{spont})} \quad (1)$$

574 where $\langle \rangle$ denotes mean, std denotes the standard deviation, r_{stim} is the average activity in response
575 to stimuli, and r_{spont} indicates the average activity over the 30-minute gray screen presentation.

576 In monkeys, we followed the definition in *Chen et al. (2022)* and calculated the SNR for each
577 site as:

$$SNR_{monkey} = \frac{\max(\langle r_{stim} \rangle) - \langle r_{spont} \rangle}{std(r_{spont})} \quad (2)$$

578 using the peak activity during the checkerboard presentation for the signal, and the average gray
579 screen neuronal activity as background (denoted as r_{spont}).

580 In mice, the split-half consistency was calculated by correlating the average activity for the 32
581 stimuli in half of the trials, randomly chosen, with the average activity in the other half of the trials,
582 followed by Spearman-Brown correction (used to correct for the division of trials by half). In mon-
583 keys, during checkerboard presentations, the split-half consistency was calculated by correlating

584 the average activity of the 16 timepoints (0–400 ms; 25 ms bins) of checkerboard presentation of
585 25 random trial repetitions with the average activity of another non-overlapping 25 random rep-
586 etitions, followed by Spearman-Brown correction. During moving bar presentations, the 40 time-
587 points (0–1s; 25 ms bins) during 25 random trial repetitions were first concatenated across the 4
588 directions (total of 160 timepoints), and then correlated to the concatenated averaged activity of
589 another nonoverlapping 25 random trial repetitions, followed by Spearman-Brown correction. For
590 all split-half consistency calculations, we randomly sampled trials 20 times.

591 **Inter-areal predictions.** Let $r_{i,t}$ be the activity of neuron or site i in area A at timepoint t , where A
592 can be L4 or L2/3 in the mouse data and V1 or V4 in the monkey data. Neuronal activity from one
593 area (e.g., mouse V1 L4 or monkey V1) was used to predict activity in the other area (e.g., mouse
594 V1 L2/3 or monkey V4) using ridge regression (**Figure 1E**). The activity of each neuron i in area A_2
595 was predicted from n_{A_1} neurons in area A_1 as follows:

$$A_2 \hat{r}_{i,t} = \sum_{j=1}^{n_{A_1}} w_{i,j} r_{j,t} + b_i \quad (3)$$

596 During fitting, we minimized the residual sum of squares (RSS), defined as:

$$RSS_i(\mathbf{w}, b_i) = \sum_{t=1}^{n_T} (A_2 \hat{r}_{i,t} - A_2 r_{i,t})^2 + \alpha \sum_{j=1}^{n_{A_1}} w_j^2 \quad (4)$$

597 where \mathbf{w} is the weight vector for predicting the activity of neuron i , n_T is the number of images/time
598 points and α controls the regularization strength (α was tuned for each dataset with an independent
599 sample and ranged from 10^3 to 10^5). Predictability for each neuron was evaluated using 10-fold
600 cross-validation across trials and quantified as squared Pearson's r between held-out $A_2 r_{i,t}$ and
601 $A_2 \hat{r}_{i,t}$, referred to as explained variance fraction (EV fraction) throughout.

602 To remove temporal auto-correlation that would inflate the apparent prediction despite cross-
603 validation, we removed training timepoints near the test timepoints closer than the decay window
604 of the activity auto-correlation (mouse: 5 s; monkey: 1.5 s). The auto-correlation decay window
605 was determined using time-series forecasting Ridge Regression (using r_t to predict r_{t+d} , where d
606 represents a delay). The delay was increased until the EV fraction approached chance.

607 To model a Poisson distribution of MUAe responses, we also fit a Poisson generalized linear
608 model (GLM) using the same predictors as above.

609 We implemented this model with scikit-learn's `PoissonRegressor` (log link; intercept b_i unpenal-
610 ized) and tuned α per dataset using an independent sample. Model evaluation followed the same
611 10-fold cross-validation and temporal exclusion procedure as ridge regression (1.5 s exclusion win-
612 dower for monkey). Predictability was quantified as the squared Pearson correlation between ob-
613 served and predicted MUAe on held-out folds (EV fraction), allowing direct comparison between
614 ridge regression and Poisson GLM (**Figure Supplement 1M**).

615 **Prediction directionality.** We compared predictability across layers in different directions (in
616 mice: L4→L2/3 vs. L2/3→L4) and also predictability across areas in different directions (in mon-
617 keys: V1→V4 vs. V4→V1) (**Figure 3**). To ensure that results were not dependent on the number
618 of neurons/sites, we randomly subsampled the number of neurons/sites of the area containing
619 the larger number of neurons/sites (L2/3 for mouse; V1 for monkeys) to match the number of
620 predictors in both directions (10 iterations per recording type, neuron count details in **Table 1** and
621 **Table 2**. Results in **Figure 3** represent the median across the 10 subsamples). To account for poten-
622 tial changes in intrinsic predictability, we ensured that the neurons from both areas were matched
623 in terms of the distribution of split-half correlation values so that the difference between individ-
624 ual area neurons/sites was less than 0.002. To assess the intrinsic predictability of neurons/sites
625 in each region, the areas were used to predict themselves, where one neuron/site in the area was
626 predicted by the remaining neurons in the same area. This “intra-areal” prediction was used to
627 normalize EV fraction to compare directionality of prediction.

628 **Stimulus types and spontaneous activity comparison.** We compared predictability for different
 629 stimulus conditions (**Figure 4, Figure 5**). To compare inter-areal prediction across stimulus types
 630 and between the presence or absence of stimuli, the number of predictors (neurons or sites) and
 631 timepoints was sub-sampled to be the same across all datasets. In monkeys, the time spent record-
 632 ing the lights-off condition was much greater than during stimulus or gray screen presentations.
 633 To account for the difference in time duration and therefore training size, we subsampled time
 634 periods to be the same across all stimulus, gray-screen, and lights-off, lights-off eyes open, and
 635 lights-off eyes closed conditions.

636 **Neuron properties.** We compared different neuronal properties with predictability measure-
 637 ments (**Figure 6**). The SNR and split-half correlation has been defined above. The absolute max
 638 pairwise correlation value of each neuron/site i in one area with all neurons in the other area was
 639 calculated as

$${}_{A2}maxcorr_i = \max_j (|\text{corr}({}_{A2}r_i, {}_{A1}r_j)|) \quad (5)$$

640 where ${}_{A2}r_i$ represents the activity of neuron/site i in area A2, which are correlated with the activity
 641 of every neuron j in area A1 (denoted as ${}_{A1}r_j$).

642 One-vs-rest self-consistency (1-vs-rest r^2). For each neuron/site i with T repeat trials, let $\mathbf{r}_i^{(t)} \in$
 643 \mathbb{R}^M be its activity vector from repetition t across all stimuli/timepoints (length M , defined below).
 644 The leave-one-out mean response over repetitions *excluding* t is

$$\bar{\mathbf{r}}_i^{(rest)} = \frac{1}{T-1} \sum_{\substack{t'=1 \\ t' \neq t}}^T \mathbf{r}_i^{(t')}. \quad (6)$$

645 Throughout, the superscript (*rest*) means “all repetitions excluding repeat t ”.

646 The one-vs-rest correlation for the held-out repetition t is

$$SC_i^{(t)} = \text{corr}(\mathbf{r}_i^{(t)}, \bar{\mathbf{r}}_i^{(rest)}), \quad (7)$$

647 We then average these correlations across all held-out repetitions:

$$\overline{SC}_i = \frac{1}{T} \sum_{t=1}^T SC_i^{(t)}. \quad (8)$$

648 In mice, $\mathbf{r}_i^{(t)}$ stacks the responses to the $S = 32$ stimuli for repetition t (each element is the mean
 649 of the two frames immediately post-onset). In monkeys, during checkerboard presentations, $\mathbf{r}_i^{(t)}$
 650 stacks the $T = 16$ timepoints (0–400 ms; 25 ms bins). In monkey moving-bar presentations, $\mathbf{r}_i^{(t)}$
 651 concatenates the $T = 160$ timepoints (4 directions \times 40 bins; 0–1 s, 25 ms bins).

652 Variance metrics. All variances were computed on gray-baseline-subtracted signals. In mice
 653 (32 gratings, K repeats), for neuron i , let $r_i^{(k,s)}$ denote the scalar response to stimulus $s \in \{1, \dots, S\}$
 654 on repeat $k \in \{1, \dots, K\}$ (repeat-averaged within the analysis window). We defined:

$$\text{Across-stimulus variance: } v_i^{\text{stim}} = \text{Var}_s(\bar{r}_i^{(s)}), \quad \text{where } \bar{r}_i^{(s)} = \frac{1}{K} \sum_{k=1}^K r_i^{(k,s)}. \quad (9)$$

655

$$\text{Trial-to-trial (within-stimulus) variance: } v_i^{\text{trial}} = \frac{1}{S} \sum_{s=1}^S \text{Var}_k(r_i^{(k,s)}). \quad (10)$$

656 In monkeys, for the checkerboard stimuli, for site i let $r_i^{(k,t)}$ denote the scalar response to time-
 657 point $t \in \{1, \dots, T\}$ on repeat $k \in \{1, \dots, K\}$

$$\text{Within-trial temporal variance: } v_i^{\text{temp}} = \text{Var}_t(\bar{r}_i^{(t)}), \quad \text{where } \bar{r}_i^{(t)} = \frac{1}{K} \sum_{k=1}^K r_i^{(k,t)}. \quad (11)$$

$$\text{Across-trial variance (per timepoint): } v_i^{\text{across}} = \frac{1}{T} \sum_{t=1}^T \text{Var}_k(r_i^{(k,t)}). \quad (12)$$

658 In monkeys, for the moving bar stimuli, the same two metrics were computed per stimulus s
659 and then averaged across the four stimuli. In addition, we defined an across-stimuli variance of
660 the time-averaged, trial-mean response:

$$v_i^{\text{stim}} = \text{Var}_s(\bar{r}_i^{(s)}), \quad \text{where } \bar{r}_i^{(s)} = \frac{1}{K} \sum_{k=1}^K \left(\frac{1}{T} \sum_{t=1}^T X_i^{(k,s)}(t) \right), \quad (13)$$

661 These variance covariates (together with SNR, split-half self-consistency, and one-vs-rest r^2 ,
662 where applicable) were included as regressors in the *within-dataset-type* linear models used to
663 obtain residual explained variance, $EV_{\text{resid}} = EV - \widehat{EV}$.

664 **EV residualization.** For each direction (V1→V4, V4→V1) and stimulus, we fit an ordinary least
665 squares (OLS) model:

$$\widehat{EV}_i = \beta_0 + \beta_1 \text{SNR}_i + \beta_2 (\text{split-half } r_i) + \beta_3 \text{Var}_{\text{time},i} + \beta_4 \text{Var}_{\text{trials},i} + \varepsilon_i \quad (14)$$

666 where \widehat{EV}_i is the predicted explained variance of each neuron/site i . Predictors were z-scored
667 within each direction or stimulus. SNR_i is its signal-to-noise ratio (defined in the *Visual Reliability*
668 section), split-half r_i is its split-half consistency (also defined above), $\text{Var}_{\text{time},i}$ and $\text{Var}_{\text{trials},i}$ are the
669 temporal and across-trial variance metrics (defined in the *Variance metrics* section), β_0, \dots, β_4 are re-
670 gression coefficients, and ε_i is the residual error term. Predictors were z-scored within each direc-
671 tion or stimulus. Residual EV was defined as $EV_{\text{resid},i} = EV_i - \widehat{EV}_i$ and used for all post-adjustment
672 comparisons (*Figure Supplement 3E-H*; *Figure Supplement 5K,O*).

673 **Dip test of unimodality.** We used Hartigan's dip test to evaluate whether the distribution of 1-vs-
674 rest squared correlation values was unimodal or not (*Figure Supplement 7A,D*). The test was per-
675 formed after restricting the analysis to highly predictable neurons/sites ($EV > 0.4$). The dip statistic
676 measures the maximum deviation between the empirical cumulative distribution function and the
677 closest unimodal distribution (*Hartigan and Hartigan, 1985*). Reported p -values correspond to the
678 probability of obtaining a dip statistic at least as large under the null hypothesis of unimodality.
679 Dip tests were computed separately for each cortical layer and mouse (Python package `dipTest`).

680 **Receptive field overlap comparisons.** In monkeys L and A, receptive field (RF) ellipses were cal-
681 culated using center and edge locations in the dataset (*Figure 6E, F*). To calculate the percentage
682 of RF overlap between the neuronal sites to be predicted and the predictor, the intersection area
683 between ellipses was retrieved using the Shapely Python package, and divided by the area of the
684 predicted site. Sites that had predictors that overlap both more than 80% and less than 10% were
685 selected to compare inter-areal predictions. To control for predictor size, 14 random predictor
686 sites from all the sites in each overlap type were subsampled (10 random samples without replace-
687 ment).

688 **Trial repeat shuffling and time offset predictions.** For the shuffled-trial experiments, we shuf-
689 fled the predictor activity across repeat trials showing the same stimulus (*Figure 7*). Thus, the
690 stimulus order remained the same. For the mouse time-offset analysis, the activity of predictor
691 neurons was time-shifted in the positive or negative direction, with 1 bin corresponding to 1 stim-
692 ulus presentation (800–900ms). For the monkey dataset, the predictor activity (400 or 1000 ms per
693 presentation, 16–50 bins of 10 ms each) was offset across time bins. We used sub-windows of 200
694 ms to avoid window-length differences that would otherwise be introduced if we shifted the entire
695 trial response.

696 **Data and code availability.** All the computational models and code for data analyses are pub-
697 licly available at this [link](#). All the data are publicly available for [mouse](#) (*Stringer et al., 2019a*), for
698 [monkeys L and A](#) (*Chen et al., 2022*), and for [monkey D](#).

Figures and Tables 699

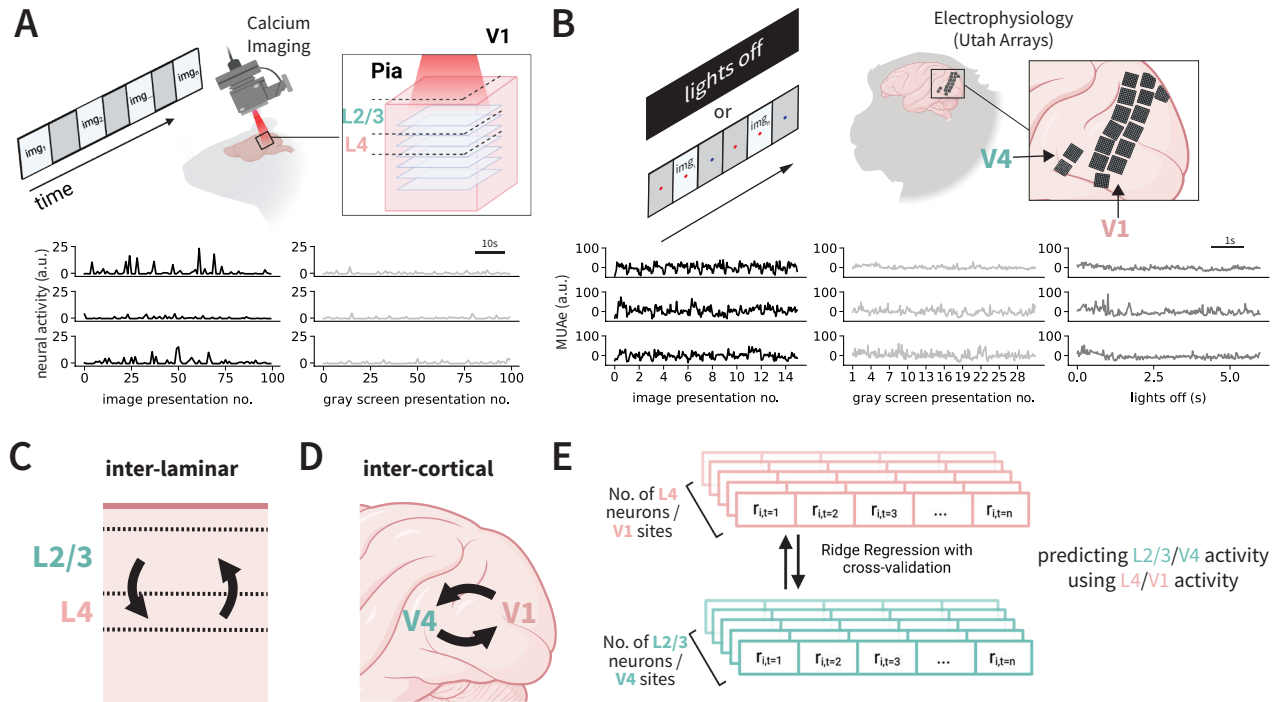


Figure 1. Predicting trial-to-trial and timepoint-to-timepoint neuronal activity between areas. **A.** Top: Experimental set-up to record two-photon Calcium imaging activity data from layers 2/3 (L2/3) and layer 4 (L4) in rodent V1 upon presentation of gratings, natural stimuli or gray screen images (represented as img_n) (Stringer et al., 2019a). Deconvolved calcium imaging traces were z-scored using baseline activity during 30 minutes of gray screen presentation before/after image presentation (Table 1). Bottom: Sample z-scored neuronal activity from 3 different neurons in response to 100 presentations of drifting gratings (left) or gray screen presentations (right). Each activity value corresponds to one image presentation, and was calculated as the average of two calcium imaging video frames (666 ms or 800 ms, see details in Methods). **B.** Top: Experimental set-up for the neuronal activity data from monkeys V1 and V4 (Chen et al., 2022). Electrophysiological activity was simultaneously recorded across 1,024 channels from 16 Utah arrays (Table 2). Bottom: Envelope multiunit spiking activity (MUAe) from 3 different sites in response to multiple presentations of a repeated 400 ms full-field checkerboard image (left, baseline mean-subtracted), 200 ms gray screen (middle), or during a lights-off condition (30 minutes total; right). Each value corresponds to aggregated MUAe activity in a 25-ms bin. **C.** Overview of inter-laminar relationships examined in mouse V1. "Lower level" layer 4 (L4) neuronal activity is used to predict "higher level" layer 2/3 (L2/3) activity and vice versa. **D.** Overview of inter-cortical relationships examined in monkeys, where lower-level V1 is used to predict higher-level V4 (blue arrow) and vice versa (red arrow). **E.** Illustration of linear ridge regression method used for inter-areal prediction. Neuronal activity in response to presentation number i (labeled " r_i ") at time t from one area (e.g., mouse V1 L2/3 or monkey V1) was used to predict activity in the other area (e.g., mouse V1 L4 or monkey V4) (Semedo et al., 2019). Predictability was evaluated using 10-fold cross-validation across presentation trials in mice, and across 25-ms timepoints in monkeys (Methods).

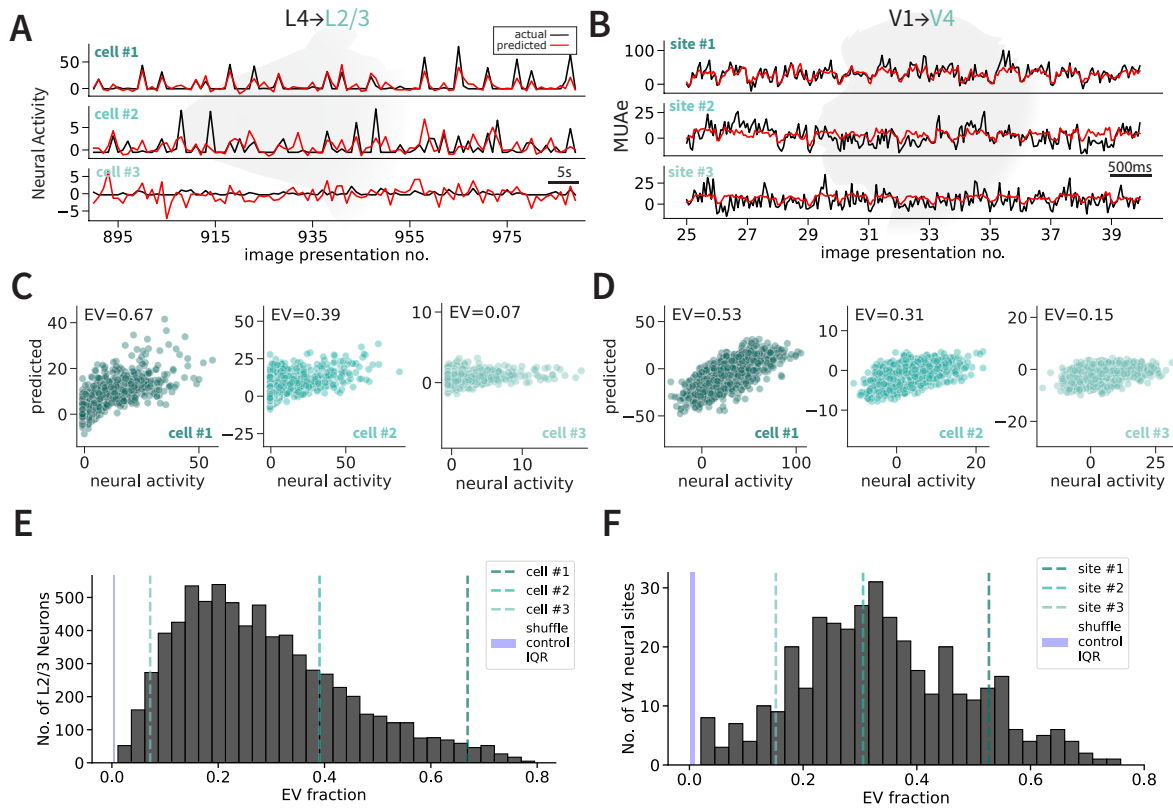


Figure 2. Lower level activity can predict higher level activity in both rodent and primate brains. **A.** Example neuronal activity (z-scored, black) in response to stimulus presentations (drifting gratings) in mouse V1 L2/3 along with regression-model predictions (red) for a typical cell (2, middle), cell in the top 10% percentile of predictability (1, top), and bottom 10% percentile (3, bottom). **B.** Same as A for monkey MUAE activity in response to a full-field checkerboard image in three V4 neuronal sites. **C.** Predicted neuronal activity versus actual neuronal activity in response to stimuli for the mouse L2/3 cells 1, 2, and 3 shown in A. Each point represents 800 ms, corresponding to a stimulus presentation. r values (top left) indicate the correlation coefficient. **D.** Same as C for monkey V4 neuronal sites 1, 2, and 3. Each point represents one 25-ms timepoint during the 400-ms presentation. **E.** Distribution of EV fraction in $L4 \rightarrow L2/3$ regressions of neural activity in response to visual stimuli in cells that were deemed visually reliable in 4 mice and 7 recording days ($n = 7,265$ neurons, Methods). Performance using 10-fold cross-validation across trials was quantified as squared Pearson's r , referred to as explained variance (EV) fraction. The three vertical lines show the 3 examples in part A, C. The blue solid shaded rectangle (here and throughout) represents the interquartile range (IQR) shuffle control performance, where the activity timepoints of one area were randomly shuffled. **F.** Distribution of EV fraction in $V1 \rightarrow V4$ regressions of neural activity in response to visual stimuli in sites deemed visually reliable in monkey L (5 recording days, 68–82 V4 sites recorded per day; $n = 378$ total site recordings).

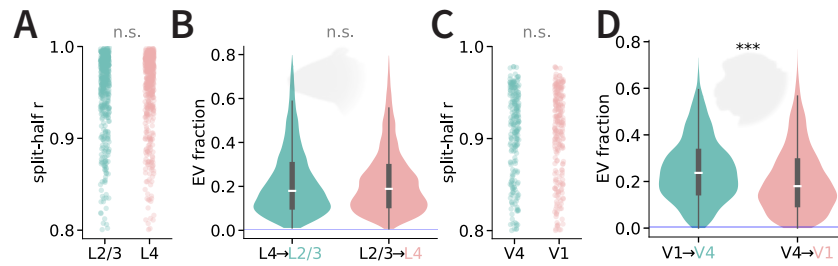


Figure 3. Asymmetry in inter-cortical predictability in monkeys but not inter-laminar predictability in mice. **A.** Split-half reliability (Methods) for the $n = 298$ neurons (per area) in mouse MP033 drifting gratings presentation recording of V1 L2/3 (green) and L4 (coral) used to perform directionality comparisons. Neurons were randomly subsampled to match the numbers and self-reliability in the two distributions. Here and throughout, asterisks indicate statistically significant differences using a hierarchical independent permutation test (10,000 permutations): * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$; “n.s.” indicates $p > 0.05$. **B.** Violin plots describing the distribution of EV fraction for L4→L2/3 (green) and L2/3→L4 (coral) predictions across all 7 stimulus recordings ($n = 1, 113$ neurons per area). Violin plots (here and throughout) represent the distribution of neuron/site values, with width representing density and inner boxplot representing the interquartile range. Whiskers of each inner box represent the data range. **C.** Example of split-half reliability for the $n = 74$ sites (per area) in monkey L checkerboard recording (date=090817) of V4 (green) and V1 (coral) used to perform directionality comparisons. **D.** Violin plots describing the distribution of EV fraction for V1→V4 (green) and V4→V1 (coral) across all 5 stimulus recordings ($n = 786$ sites recordings per area).

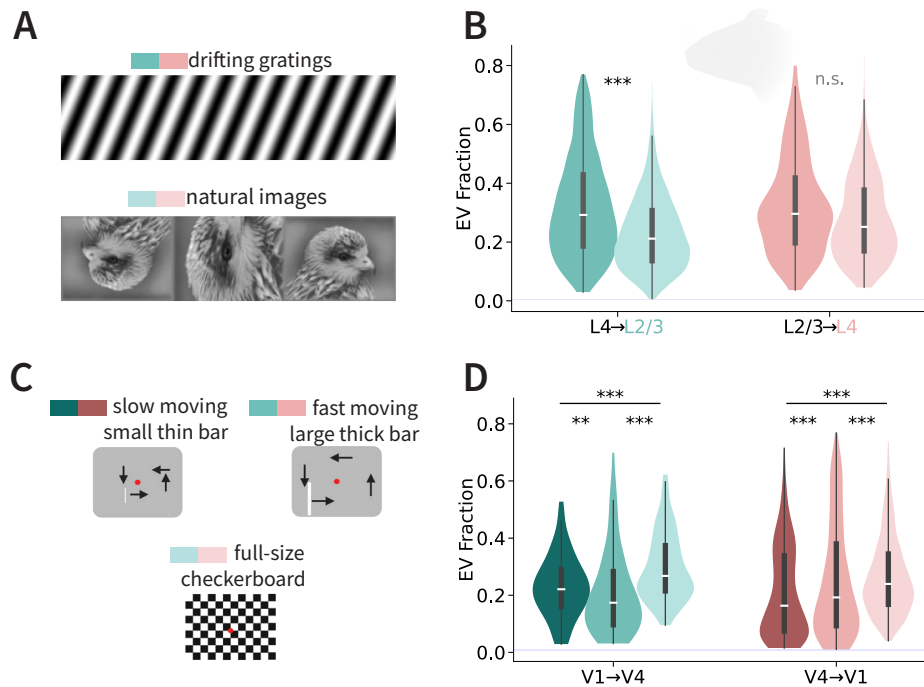


Figure 4. Stimulus type influences neuronal predictability. **A.** Illustration of the two types of stimuli (drifting gratings and static natural images) presented to the mouse during calcium imaging. **B.** Across-layer predictability in mouse V1 for each stimulus type (dark: drifting gratings, light: natural images) and prediction direction. **C.** Illustration of the three types of stimuli presented to the monkeys (*Chen et al., 2022*). The slow-moving, small, thin bar moved near the fixation point for 1 s in each of the four directions, while the fast-moving, large, thick bar moved towards the edges of the screen for 1 s in each of the four directions. The full-field checkerboard image was presented repeatedly (400 ms each presentation). **D.** Across-area predictability for each stimulus type (dark: slow bars, medium: fast bars, light: checkerboard) and direction.

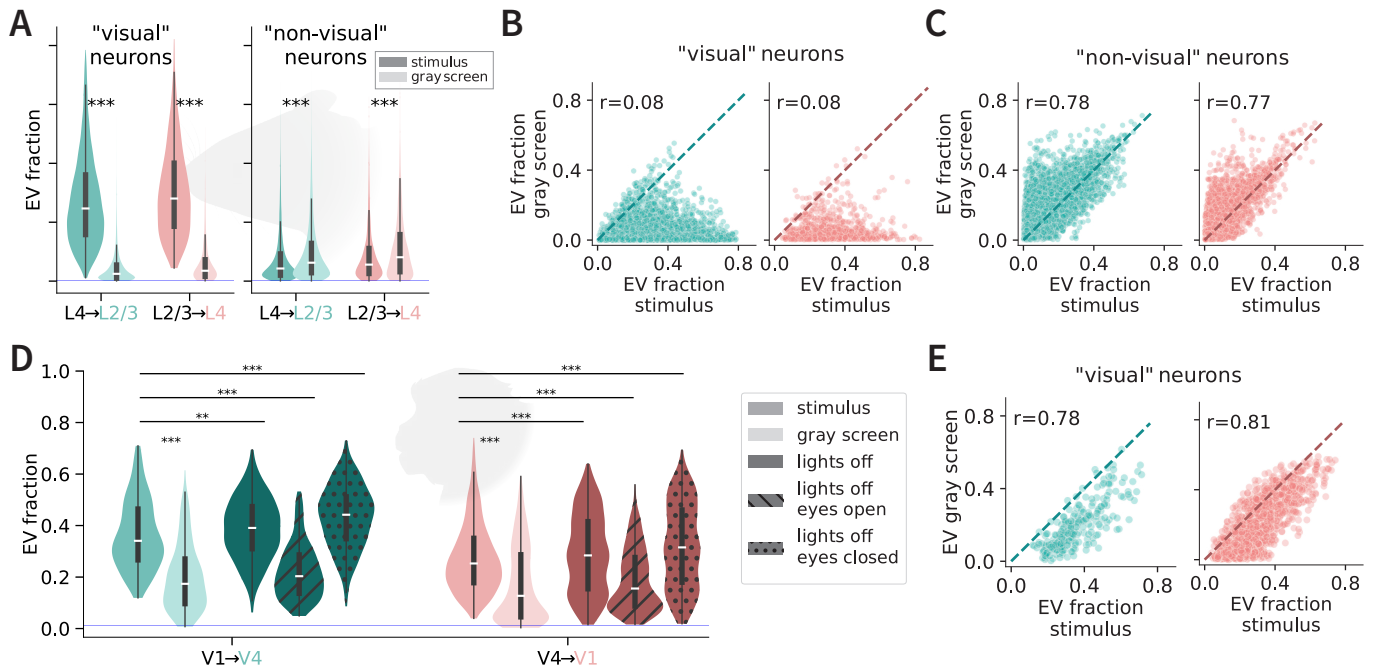


Figure 5. Spontaneous activity can also be predicted. **A.** EV fraction of neuronal activity in response to stimulus presentation (dark violins) or gray screen presentation (light violins) for neurons deemed visually (left) or non-visually (right) reliable (Methods). **B.** Correlation between EV in responses to gray screen (y-axis) versus stimulus presentation (x-axis) in mouse V1 visually reliable neurons (L4→L2/3:left, green; L2/3→L4: right, coral). The diagonal line represents the line of equality ($y=x$). r is the Pearson's r coefficient. **C.** Same as **B**, but for non-visually reliable neurons. **D.** EV during stimulus presentations (checkerboard image, green), gray screen presentations (light green), or during lights off (dark green). The lights-off condition is further separated into periods when the eyes were open or closed. All lights-off conditions were sub-sampled (10 permutations) to contain similar training lengths as the stimulus and gray screen presentation recordings. **E.** Correlation between EV in responses to gray screen (y-axis) versus stimulus presentation (x-axis) in monkey visually reliable neurons (V1→V4:left, green; V4→V1: right, coral). The diagonal line represents the line of equality ($y=x$). r is the Pearson's r coefficient. All recorded sites were pulled from the 3 recording days of checkerboard presentations.

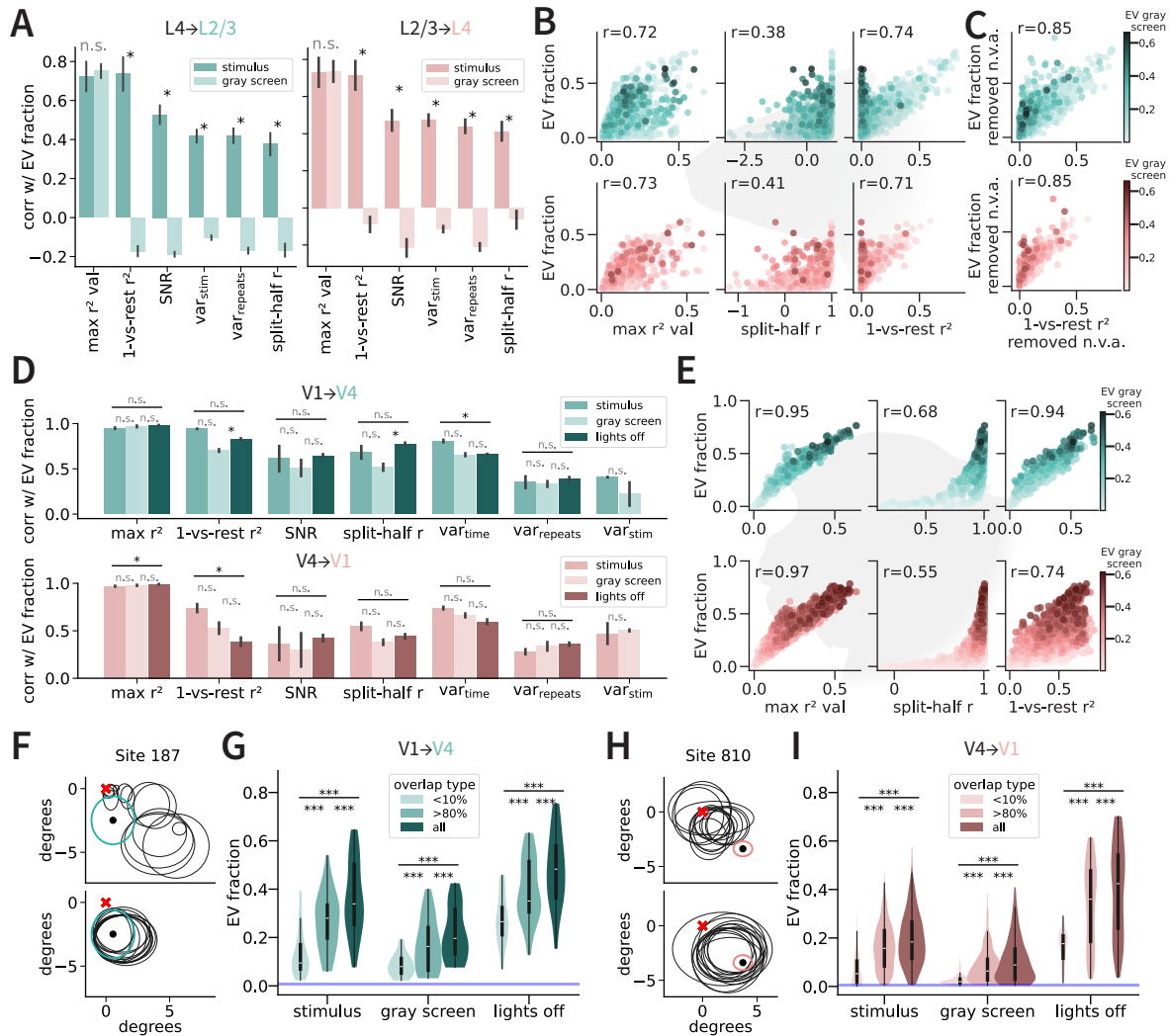


Figure 6. neuronal predictability depends on SNR, stimulus response variance, and receptive field overlap. **A.** Correlation between different neuronal properties with the predictability of L2/3 (green) and L4 (coral) neuronal responses during the presence (dark color) or absence (light color) of visual stimulus. Neuronal properties measured in mouse V1 include the correlation value of the most correlated pair to each cell (max correlation value, squared), a modified metric of self-consistency (one-vs-rest correlation, squared), SNR, variance in the neuronal activity in response to different stimuli, variance in the neuronal activity across repetitions, and the traditional metric of self-consistency (split-half correlation r) (Methods). **B.** Relationship between three neuronal properties and their predictability in a randomly chosen sub-sample of neurons ($n = 4,000$) for mouse L2/3 (green) and L4 (coral) neuronal responses from both drifting gratings and natural images conditions (combined). Hue represents the degree of predictability for the same neurons during the 30 minutes of gray screen presentation (see color map on bottom right). **C.** 1-vs-rest square correlation relationship with predictability after projecting out dimensions of "non-visual" activity (using gray screen activity (Stringer et al., 2019a)). **D.** Correlation between different neuronal properties with the predictability of monkey L V4 (green) and V1 (coral) neuronal site recordings during the full-field checkerboard presentation (dark color), gray screen presentation (light color), and lights-off condition (darkest color; solid, hatch lines, and hatch dots). Neuronal properties measured in monkey visual cortex include the max correlation squared value, one-vs-rest squared correlation, SNR, variance across different stimuli (moving bars dataset only), variance across time (within-trial repeat), variance across repeats (within timepoint), and split-half correlation r . **E.** Same as B for monkey L V1 and V4 neuronal sites. **F.** Top: Receptive fields of one sample V4 neuronal site (green circle, array 2 electrode 187) and 14 randomly selected V1 neuronal sites as predictors (black circles), constrained on sites that share less than 10% receptive field overlap with the V4 site. Bottom: Receptive fields of the same neuronal site 187 and 14 randomly selected V1 neuronal sites used as predictors, constrained on sites that share at least 80% receptive field overlap with the V4 site. **G.** Differences in predictability of V4 neural activity ($n = 110$ site recordings) in terms of 14 V1 predictor sites with less than 10% RF overlap (light green), 14 predictor sites with at least 80% RF overlap (green), and all predictors (dark green). Predictions were computed for recordings in response to the stimulus presentation (sliding bars and full-field checkerboard images), gray screen presentation, and lights off. **H.** Bottom and top left: Same as F but for monkey L sample V1 site 810. **I.** Same as D, but for V1 ($n = 970$ site recordings).

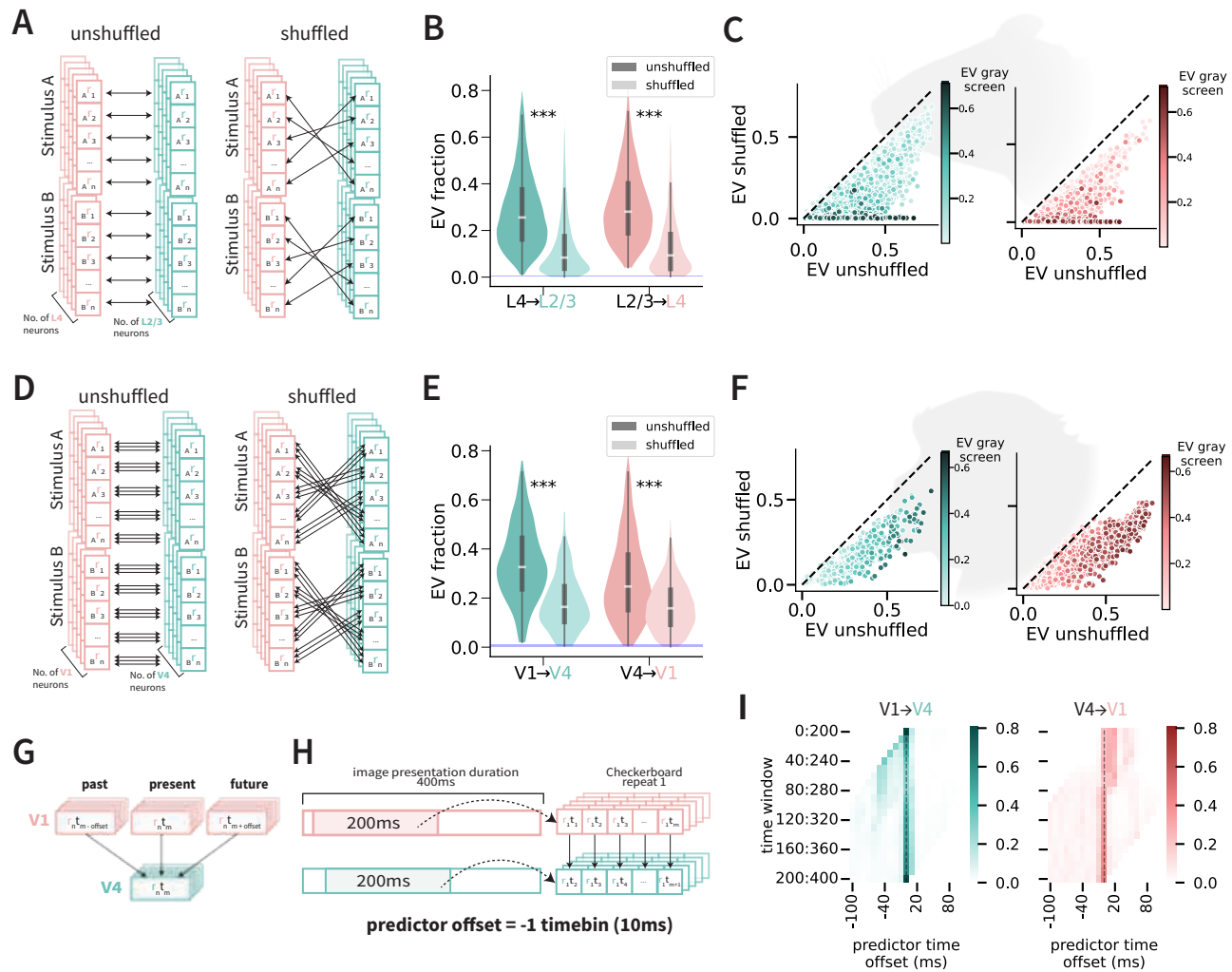


Figure 7. Predicting neuronal activity across time reveals shared stimulus- and non-stimulus driven information in both mouse and monkey visual cortex, along with latency and non-latency specific correlations in monkey V1/V4. **A.** Shuffled-trial-repeat experiment set-up for comparing unshuffled vs. shuffled prediction in mouse V1 L2/3 and L4. Neuronal activity in response to stimulus repeats was shuffled within their respective image. **B.** EV fraction for unshuffled (dark) and shuffled (light) trials in the L4 → L2/3 (green) and L2/3 → L4 (coral) directions. **C.** Relationship between shuffled (y-axis) and unshuffled (x-axis) trial repeat EVs in the mouse L4 → L2/3 (left, green) and L2/3 → L4 (right, coral) directions. Hue represents EV fraction during gray screen activity. **D.** Shuffled-trial-repeat experiment set-up comparing unshuffled vs. shuffled prediction in monkey data. Neuronal activity (including all timepoints) in response to stimulus repeats was shuffled within their respective image. Since the checkerboard presentation was only one stimulus, the experiment visualization only applies to the “Stimulus A” portion. **E.** Same as **B** for monkey L V1 → V4 (green) and V4 → V1 (coral). **F.** Same as **C** for monkey V1 → V4 (green) and V4 → V1 (coral). **G.** Illustration of time offsets applied to monkey neuronal activity for inter-areal predictions. Instead of neuronal activity prediction between areas being done on simultaneous activity (middle coral and bottom green box), the V4 neuronal activity (green) at time t_m was predicted using V1 neuronal activity (coral) at time $t_{m \pm \text{offset}}$. *offset* represents 1–8 timebins (25 ms per timebin) before (if negative; left coral box) or after (if positive; right coral box) time t_m . Time offset experiment was done in both prediction directions (V1 → V4 and V4 → V1). **H.** Experimental set-up example for predicting neuronal activity in V4 using V1 neuronal activity from 10 ms prior to V4 activity. Neural activity is in response to a repeated checkerboard image. A 200 ms section of the cortical area was used to represent the image presentation response, and was offset -1 timebin (10 ms) to predict a 200 ms target cortical area. During the prediction experiments, the 200 ms window was slid across the entire duration of the stimulus. **I.** Time offset prediction results across both V1 → V4 (left, green) and V4 → V1 (right, coral) prediction directions. Each square corresponds to the fraction of neuronal sites whose neural activity were best predicted during that offset period and time window.

Table 1. Mouse neuron counts used for inter-layer prediction and analyses. A total of 7 recordings were used to perform prediction experiments. Each row corresponds to a recording day, containing the dataset recording type (Mouse Dataset), total number of neurons, and visually reliable neurons (see *Methods*). Fourth column: In the directionality prediction experiments, the area containing more neurons (L2/3) was further subsampled to match the number of L4 neurons. The dataset recording type names contain either “ori32” or “nating32”, in addition to the mouse name (MP0-). “nating32” represents the dataset of the 32 natural image presentations. “ori32” represents the dataset of the 32 drifting gratings.

Mouse Dataset	Layer 2/3			Layer 4	
	Total	visually reliable	Subsampled (directionality)	Total	visually reliable
nating32 MP031	6615	1248	219	2367	219
nating32 MP032	7980	1549	96	1441	96
nating32 MP033	6646	1467	164	2010	164
ori32 MP027	6264	1029	211	2346	211
ori32 MP031	5423	455	78	1382	78
ori32 MP032	5420	274	47	923	47
ori32 MP033	5277	1243	298	1588	298
Total	43625	7265	1113	12057	1113

Table 2. Monkey site counts used for inter-cortical prediction and analyses. First column: First letter denotes the monkey name, followed by the date, followed by the dataset type acquired during the session (C: Checkerboard presentations, G: Gray screen, L: Lights-off condition, LB: Large, thick moving bars, SB: Small, thin moving bars). Fourth column: In the directionality prediction experiments, the area containing more sites was further subsampled to match the number of V4 sites.

Monkey Datasets	V1			V4	
	Total	visually reliable	Subsampled (directionality)	Total	visually reliable
L 090817 (C,G,L)	627	555	73	96	73
L 100817 (C,G,L)	688	587	83	115	83
L 250717 (C,G,L)	645	537	72	86	72
L 260617 (LB,G)	645	592	82	86	82
L 280617 (SB,G)	645	518	68	86	68
A 041018 (C,G)	571	357	44	76	44
A 280818 (LB,G)	571	381	58	76	58
A 290818 (SB,G)	571	251	30	76	30
D 250225 (C,G,L)	16	8	-	16	7
D 260225 (C,G,L)	16	10	-	16	10
Total	4995	3796	510	729	527

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707 R01 NS113890 to SS.

708 References

- 709 **Andermann ML**, Kerlin AM, Roumis DK, Glickfeld LL, Reid RC. Functional Specialization of Mouse Higher Visual
710 Cortical Areas. *Neuron*. 2011 Dec; 72(6):1025–1039. [https://www.cell.com/neuron/abstract/S0896-6273\(11\)](https://www.cell.com/neuron/abstract/S0896-6273(11)01012-9)
711 [01012-9](https://doi.org/10.1016/j.neuron.2011.11.013), doi: 10.1016/j.neuron.2011.11.013, publisher: Elsevier.
- 712 **Avitan L**, Stringer C. Not so spontaneous: Multi-dimensional representations of behaviors and context in
713 sensory areas. *Neuron*. 2022 Oct; 110(19):3064–3075. [https://www.sciencedirect.com/science/article/pii/](https://www.sciencedirect.com/science/article/pii/S0896627322005888)
714 [S0896627322005888](https://doi.org/10.1016/j.neuron.2022.06.019), doi: 10.1016/j.neuron.2022.06.019.
- 715 **Bastos AM**, Vezoli J, Bosman CA, Schoffelen JM, Oostenveld R, Dowdall JR, De Weerd P, Kennedy H, Fries
716 P. Visual Areas Exert Feedforward and Feedback Influences through Distinct Frequency Channels. *Neu-*
717 *ron*. 2015 Jan; 85(2):390–401. <https://www.sciencedirect.com/science/article/pii/S089662731401099X>, doi:
718 [10.1016/j.neuron.2014.12.018](https://doi.org/10.1016/j.neuron.2014.12.018).
- 719 **Chacron MJ**, Longtin A, Maler L. The effects of spontaneous activity, background noise, and the stimulus
720 ensemble on information transfer in neurons. *Network (Bristol, England)*. 2003 Nov; 14(4):803–824.
- 721 **Chen X**, Morales-Gregorio A, Sprenger J, Kleinjohann A, Sridhar S, van Albada SJ, Grün S, Roelfsema PR. 1024-
722 channel electrophysiological recordings in macaque V1 and V4 during resting state. *Scientific Data*. 2022 Mar;
723 9(1):77. <https://www.nature.com/articles/s41597-022-01180-1>, doi: 10.1038/s41597-022-01180-1, number: 1
724 Publisher: Nature Publishing Group.
- 725 **Connor CE**, Brincat SL, Pasupathy A. Transformation of shape information in the ventral pathway. *Current*
726 *Opinion in Neurobiology*. 2007 Apr; 17(2):140–147. doi: 10.1016/j.conb.2007.03.002.
- 727 **Consortium M**, Bae JA, Baptiste M, Bodor AL, Brittain D, Buchanan J, Bumbarger DJ, Castro MA, Celii B, Cobos E,
728 Collman F, Costa NMd, Dorkenwald S, Elabbady L, Fahey PG, Fliss T, Froudarakis E, Gager J, Gamlin C, Halageri
729 A, et al. Functional connectomics spanning multiple areas of mouse visual cortex. *bioRxiv*; 2021.
- 730 **Dadarlat MC**, Stryker MP. Locomotion Enhances Neural Encoding of Visual Stimuli in Mouse V1. *The Jour-*
731 *nal of Neuroscience: The Official Journal of the Society for Neuroscience*. 2017 Apr; 37(14):3764–3775. doi:
732 [10.1523/JNEUROSCI.2728-16.2017](https://doi.org/10.1523/JNEUROSCI.2728-16.2017).
- 733 **Douglas RJ**, Martin KAC. Neuronal circuits of the neocortex. *Annual Review of Neuroscience*. 2004; 27:419–451.
734 doi: 10.1146/annurev.neuro.27.070203.144152.
- 735 **Felleman DJ**, Van Essen DC. Distributed hierarchical processing in the primate cerebral cortex. *Cerebral Cortex*
736 (New York, NY: 1991). 1991; 1(1):1–47. doi: 10.1093/cercor/1.1.1-a.
- 737 **Gazzaley A**, Rissman J, Cooney J, Rutman A, Seibert T, Clapp W, D'Esposito M. Functional Interactions be-
738 tween Prefrontal and Visual Association Cortex Contribute to Top-Down Modulation of Visual Processing.
739 *Cerebral Cortex*. 2007 Sep; 17(suppl_1):i125–i135. <https://doi.org/10.1093/cercor/bhm113>, doi: 10.1093/cer-
740 cor/bhm113.
- 741 **Gilbert CD**, Li W. Top-down influences on visual processing. *Nature Reviews Neuroscience*. 2013 May;
742 14(5):350–363. <https://www.nature.com/articles/nrn3476>, doi: 10.1038/nrn3476, number: 5 Publisher: Nature
743 Publishing Group.
- 744 **Gokcen E**, Jasper AI, Semedo JD, Zandvakili A, Kohn A, Machens CK, Yu BM. Disentangling the flow of signals
745 between populations of neurons. *Nature Computational Science*. 2022 Aug; 2(8):512–525. [https://www.](https://www.nature.com/articles/s43588-022-00282-5)
746 [nature.com/articles/s43588-022-00282-5](https://doi.org/10.1038/s43588-022-00282-5), doi: 10.1038/s43588-022-00282-5, number: 8 Publisher: Nature
747 Publishing Group.

- 748 **Hartigan JA**, Hartigan PM. The Dip Test of Unimodality. *The Annals of Statistics*. 1985 Mar; 13(1):70–
749 84. [https://projecteuclid.org/journals/annals-of-statistics/volume-13/issue-1/The-Dip-Test-of-Unimodality/10.](https://projecteuclid.org/journals/annals-of-statistics/volume-13/issue-1/The-Dip-Test-of-Unimodality/10.1214/aos/1176346577.full)
750 [1214/aos/1176346577.full](https://projecteuclid.org/journals/annals-of-statistics/volume-13/issue-1/The-Dip-Test-of-Unimodality/10.1214/aos/1176346577.full), doi: 10.1214/aos/1176346577, publisher: Institute of Mathematical Statistics.
- 751 **Hohaia W**, Saurels BW, Johnston A, Yarrow K, Arnold DH. Occipital alpha-band brain waves when the eyes are
752 closed are shaped by ongoing visual processes. *Scientific Reports*. 2022 Jan; 12:1194. [https://www.ncbi.nlm.](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8786963/)
753 [nih.gov/pmc/articles/PMC8786963/](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8786963/), doi: 10.1038/s41598-022-05289-6.
- 754 **Hsu A**, Borst A, Theunissen FE. Quantifying variability in neural responses and its application for the val-
755 idation of model predictions. *Network: Computation in Neural Systems*. 2004 Jan; 15(2):91–109. https://doi.org/10.1088/0954-898X_15_2_002, doi: 10.1088/0954-898X_15_2_002, publisher: Taylor & Francis
756 [_eprint: https://doi.org/10.1088/0954-898X_15_2_002](https://doi.org/10.1088/0954-898X_15_2_002).
757
- 758 **Hubel DH**, Wiesel TN. Receptive fields, binocular interaction and functional architecture in the
759 cat's visual cortex. *The Journal of Physiology*. 1962; 160(1):106–154. [https://onlinelibrary.](https://onlinelibrary.wiley.com/doi/abs/10.1113/jphysiol.1962.sp006837)
760 [wiley.com/doi/abs/10.1113/jphysiol.1962.sp006837](https://onlinelibrary.wiley.com/doi/abs/10.1113/jphysiol.1962.sp006837), doi: 10.1113/jphysiol.1962.sp006837, _eprint:
761 <https://onlinelibrary.wiley.com/doi/pdf/10.1113/jphysiol.1962.sp006837>.
- 762 **Ibrahim LA**, Schuman B, Bandler R, Rudy B, Fishell G. Mining the jewels of the cortex's crowning mys-
763 tery. *Current Opinion in Neurobiology*. 2020 Aug; 63:154–161. [https://linkinghub.elsevier.com/retrieve/pii/](https://linkinghub.elsevier.com/retrieve/pii/S0959438820300842)
764 [S0959438820300842](https://linkinghub.elsevier.com/retrieve/pii/S0959438820300842), doi: 10.1016/j.conb.2020.04.005.
- 765 **Jiang X**, Shen S, Cadwell CR, Berens P, Sinz F, Ecker AS, Patel S, Tolias AS. Principles of connectivity among
766 morphologically defined cell types in adult neocortex. *Science*. 2015 Nov; 350(6264):aac9462. [https://www.](https://www.science.org/doi/10.1126/science.aac9462)
767 [science.org/doi/10.1126/science.aac9462](https://www.science.org/doi/10.1126/science.aac9462), doi: 10.1126/science.aac9462, publisher: American Association for
768 the Advancement of Science.
- 769 **van Kerkoerle T**, Self MW, Dagnino B, Gariel-Mathis MA, Poort J, van der Togt C, Roelfsema PR. Alpha and
770 gamma oscillations characterize feedback and feedforward processing in monkey visual cortex. *Proceedings*
771 *of the National Academy of Sciences*. 2014 Oct; 111(40):14332–14341. [https://www.pnas.org/doi/full/10.](https://www.pnas.org/doi/full/10.1073/pnas.1402773111)
772 [1073/pnas.1402773111](https://www.pnas.org/doi/full/10.1073/pnas.1402773111), doi: 10.1073/pnas.1402773111, publisher: Proceedings of the National Academy of
773 Sciences.
- 774 **Kreiman G**. *Biological and Computer Vision*. Cambridge: Cambridge University Press; 2021. [https://](https://www.cambridge.org/core/books/biological-and-computer-vision/BB7E68A69AFE7A322F68F3C4A297F3CF)
775 www.cambridge.org/core/books/biological-and-computer-vision/BB7E68A69AFE7A322F68F3C4A297F3CF, doi:
776 10.1017/9781108649995.
- 777 **Lee WCA**, Bonin V, Reed M, Graham BJ, Hood G, Glatfelder K, Reid RC. Anatomy and function of an excita-
778 tory network in the visual cortex. *Nature*. 2016 Apr; 532(7599):370–374. [https://www.nature.com/articles/](https://www.nature.com/articles/nature17192)
779 [nature17192](https://www.nature.com/articles/nature17192), doi: 10.1038/nature17192, number: 7599 Publisher: Nature Publishing Group.
- 780 **Markov NT**, Ercsey-Ravasz MM, Ribeiro Gomes AR, Lamy C, Magrou L, Vezoli J, Misery P, Falchier A, Quilodran
781 R, Gariel MA, Sallet J, Gamanut R, Huissoud C, Clavagnier S, Giroud P, Sappey-Marinié D, Barone P, Dehay C,
782 Toroczkai Z, Knoblauch K, et al. A weighted and directed interareal connectivity matrix for macaque cerebral
783 cortex. *Cerebral Cortex (New York, NY)*. 2014 Jan; 24(1):17–36. doi: 10.1093/cercor/bhs270.
- 784 **Morales-Gregorio A**, Kurth AC, Ito J, Kleinjohann A, Barthélemy FV, Brochier T, Grün S, van Albada SJ.
785 Neural manifolds in V1 change with top-down signals from V4 targeting the foveal region. *Cell Re-*
786 *ports*. 2024 Jul; 43(7):114371. <https://www.sciencedirect.com/science/article/pii/S2211124724006995>, doi:
787 [10.1016/j.celrep.2024.114371](https://www.sciencedirect.com/science/article/pii/S2211124724006995).
- 788 **Niell CM**, Stryker MP. Highly Selective Receptive Fields in Mouse Visual Cortex. *The Journal of Neuroscience*.
789 2008 Jul; 28(30):7520. <https://pmc.ncbi.nlm.nih.gov/articles/PMC3040721/>, doi: 10.1523/JNEUROSCI.0623-
790 [08.2008](https://pmc.ncbi.nlm.nih.gov/articles/PMC3040721/).
- 791 **Niell CM**, Stryker MP. Modulation of visual responses by behavioral state in mouse visual cor-
792 tex. *Neuron*. 2010 Feb; 65(4):472–479. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3184003/>, doi:
793 [10.1016/j.neuron.2010.01.033](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3184003/).
- 794 **Okazaki Y**, Abrahamyan A, Stevens CJ, Ioannides AA. The timing of face selectivity and attentional modulation
795 in visual processing. *Neuroscience*. 2008 Apr; 152(4):1130–1144. [https://www.sciencedirect.com/science/](https://www.sciencedirect.com/science/article/pii/S0306452208001541)
796 [article/pii/S0306452208001541](https://www.sciencedirect.com/science/article/pii/S0306452208001541), doi: 10.1016/j.neuroscience.2008.01.056.
- 797 **Papadopouli M**, Smyrnakis I, Koniotakis E, Savaglio MA, Brozi C, Psilou E, Palagina G, Smirnakis SM. Brain
798 orchestra under spontaneous conditions: Identifying communication modules from the functional archi-
799 tecture of area V1. *bioRxiv*. 2024 Mar; <https://www.biorxiv.org/content/10.1101/2024.02.29.582364v1>, doi:
800 [10.1101/2024.02.29.582364](https://www.biorxiv.org/content/10.1101/2024.02.29.582364), pages: 2024.02.29.582364 Section: New Results.

- 801 **Park J**, Papoutsi A, Ash RT, Marin MA, Poirazi P, Smirnakis SM. Contribution of apical and basal dendrites to
802 orientation encoding in mouse V1 L2/3 pyramidal neurons. *Nature Communications*. 2019 Nov; 10(1):5372.
803 <https://www.nature.com/articles/s41467-019-13029-0>, doi: 10.1038/s41467-019-13029-0, publisher: Nature
804 Publishing Group.
- 805 **Pasupathy A**, Popovkina DV, Kim T. Visual Functions of Primate Area V4. *Annual Review of Vision Science*. 2020;
806 6(1):363–385. <https://doi.org/10.1146/annurev-vision-030320-041306>, doi: 10.1146/annurev-vision-030320-
807 041306, _eprint: <https://doi.org/10.1146/annurev-vision-030320-041306>.
- 808 **Petousakis KE**, Park J, Papoutsi A, Smirnakis S, Poirazi P. Modeling apical and basal tree contribution to ori-
809 entation selectivity in a mouse primary visual cortex layer 2/3 pyramidal cell. *eLife*. 2023 Dec; 12:e91627.
810 <https://elifesciences.org/articles/91627>, doi: 10.7554/eLife.91627.
- 811 **Polack PO**, Friedman J, Golshani P. Cellular mechanisms of brain state-dependent gain modulation in visual
812 cortex. *Nature Neuroscience*. 2013 Sep; 16(9):1331–1339. doi: 10.1038/nn.3464.
- 813 **Reynolds JH**, Chelazzi L. Attentional modulation of visual processing. *Annual Review of Neuroscience*. 2004;
814 27:611–647. doi: 10.1146/annurev-neuro.26.041002.131039.
- 815 **Ringach DL**. Spontaneous and driven cortical activity: implications for computation. *Current Opinion in*
816 *Neurobiology*. 2009 Aug; 19(4):439–444. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3319344/>, doi:
817 10.1016/j.conb.2009.07.005.
- 818 **Salinas E**, Sejnowski TJ. Correlated neuronal activity and the flow of neural information. *Nature Reviews Neu-*
819 *roscience*. 2001 Aug; 2(8):539–550. <https://www.nature.com/articles/35086012>, doi: 10.1038/35086012, pub-
820 lisher: Nature Publishing Group.
- 821 **Schmolesky MT**, Wang Y, Hanes DP, Thompson KG, Leutgeb S, Schall JD, Leventhal AG. Signal Timing Across the
822 Macaque Visual System. *Journal of Neurophysiology*. 1998 Jun; 79(6):3272–3278. [https://journals.physiology.](https://journals.physiology.org/doi/full/10.1152/jn.1998.79.6.3272)
823 [org/doi/full/10.1152/jn.1998.79.6.3272](https://journals.physiology.org/doi/full/10.1152/jn.1998.79.6.3272), doi: 10.1152/jn.1998.79.6.3272, publisher: American Physiological
824 Society.
- 825 **Schuman B**, Dellal S, Prönneke A, Machold R, Rudy B. Neocortical Layer 1: An Elegant Solution to Top-Down and
826 Bottom-Up Integration. *Annual Review of Neuroscience*. 2021 Jul; 44(1):221–252. [https://www.annualreviews.](https://www.annualreviews.org/doi/10.1146/annurev-neuro-100520-012117)
827 [org/doi/10.1146/annurev-neuro-100520-012117](https://www.annualreviews.org/doi/10.1146/annurev-neuro-100520-012117), doi: 10.1146/annurev-neuro-100520-012117.
- 828 **Semedo JD**, Jasper AI, Zandvakili A, Krishna A, Aschner A, Machens CK, Kohn A, Yu BM. Feedforward and feed-
829 back interactions between visual cortical areas use different population activity patterns. *Nature Communi-*
830 *cations*. 2022 Mar; 13(1):1099. <https://www.nature.com/articles/s41467-022-28552-w>, doi: 10.1038/s41467-
831 022-28552-w, number: 1 Publisher: Nature Publishing Group.
- 832 **Semedo JD**, Zandvakili A, Machens CK, Yu BM, Kohn A. Cortical Areas Interact through a Communication Sub-
833 space. *Neuron*. 2019 Apr; 102(1):249–259.e4. <https://linkinghub.elsevier.com/retrieve/pii/S0896627319300534>,
834 doi: 10.1016/j.neuron.2019.01.026.
- 835 **Serre T**, Oliva A, Poggio T. A feedforward architecture accounts for rapid categorization. *Proceedings of*
836 *the National Academy of Sciences*. 2007 Apr; 104(15):6424–6429. [https://www.pnas.org/doi/10.1073/pnas.](https://www.pnas.org/doi/10.1073/pnas.0700622104)
837 [0700622104](https://www.pnas.org/doi/10.1073/pnas.0700622104), doi: 10.1073/pnas.0700622104, publisher: Proceedings of the National Academy of Sciences.
- 838 **Serre T**, Wolf L, Bileschi S, Riesenhuber M, Poggio T. Robust Object Recognition with Cortex-Like Mechanisms.
839 *IEEE Transactions on Pattern Analysis and Machine Intelligence*. 2007 Mar; 29(3):411–426. [https://ieeexplore.](https://ieeexplore.ieee.org/document/4069258)
840 [ieee.org/document/4069258](https://ieeexplore.ieee.org/document/4069258), doi: 10.1109/TPAMI.2007.56, conference Name: IEEE Transactions on Pattern
841 Analysis and Machine Intelligence.
- 842 **Shen S**, Jiang X, Scala F, Fu J, Fahey P, Kobak D, Tan Z, Reimer J, Sinz F, Tolias AS. Distinct organization of two
843 cortico-cortical feedback pathways. *bioRxiv*; 2020.
- 844 **Stringer C**, Recordings of ten thousand neurons in visual cortex during spontaneous behaviors. *Janelia Re-*
845 *search Campus*; 2018. [https://figshare.com/articles/dataset/Recordings_of_ten_thousand_neurons_in_](https://figshare.com/articles/dataset/Recordings_of_ten_thousand_neurons_in_visual_cortex_during_spontaneous_behaviors/6163622/6)
846 [visual_cortex_during_spontaneous_behaviors/6163622/6](https://figshare.com/articles/dataset/Recordings_of_ten_thousand_neurons_in_visual_cortex_during_spontaneous_behaviors/6163622/6), doi: 10.25378/janelia.6163622.v6.
- 847 **Stringer C**, Michaelos M, Tsybouski D, Lindo SE, Pachitariu M. High-precision coding in visual cortex. *Cell*.
848 2021 May; 184(10):2767–2778.e15. <https://www.sciencedirect.com/science/article/pii/S0092867421003731>, doi:
849 10.1016/j.cell.2021.03.042.

- 850 **Stringer C**, Pachitariu M, Steinmetz N, Carandini M, Harris KD. High-dimensional geometry of population re-
851 sponses in visual cortex. *Nature*. 2019 Jun; 571(7765):361–365. [https://www.ncbi.nlm.nih.gov/pmc/articles/](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6642054/)
852 [PMC6642054/](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6642054/), doi: 10.1038/s41586-019-1346-5.
- 853 **Stringer C**, Pachitariu M, Steinmetz N, Reddy CB, Carandini M, Harris KD. Spontaneous behaviors drive mul-
854 tidimensional, brainwide activity. *Science*. 2019 Apr; 364(6437):eaav7893. [https://www.science.org/doi/10.](https://www.science.org/doi/10.1126/science.aav7893)
855 [1126/science.aav7893](https://www.science.org/doi/10.1126/science.aav7893), doi: [10.1126/science.aav7893](https://www.science.org/doi/10.1126/science.aav7893), publisher: American Association for the Advancement
856 of Science.
- 857 **Talluri BC**, Kang I, Lazere A, Quinn KR, Kaliss N, Yates JL, Butts DA, Nienborg H. Activity in primate visual cortex
858 is minimally driven by spontaneous movements. *Nature Neuroscience*. 2023 Nov; 26(11):1953–1959. [https:](https://www.nature.com/articles/s41593-023-01459-5)
859 [://www.nature.com/articles/s41593-023-01459-5](https://www.nature.com/articles/s41593-023-01459-5), doi: 10.1038/s41593-023-01459-5, number: 11 Publisher:
860 Nature Publishing Group.
- 861 **Tang Y**, Gervais C, Moffitt R, Nareddula S, Zimmermann M, Nadew YY, Quinn CJ, Saldarriaga V, Edens P,
862 Chubykin AA. Visual experience induces 4–8 Hz synchrony between V1 and higher-order visual areas. *Cell*
863 *Reports*. 2023 Dec; 42(12):113482. <https://www.sciencedirect.com/science/article/pii/S2211124723014948>, doi:
864 [10.1016/j.celrep.2023.113482](https://www.sciencedirect.com/science/article/pii/S2211124723014948).
- 865 **Wang Q**, Burkhalter A. Area map of mouse visual cortex. *Journal of Comparative Neurology*. 2007;
866 502(3):339–357. <https://onlinelibrary.wiley.com/doi/abs/10.1002/cne.21286>, doi: [10.1002/cne.21286](https://onlinelibrary.wiley.com/doi/abs/10.1002/cne.21286), eprint:
867 <https://onlinelibrary.wiley.com/doi/pdf/10.1002/cne.21286>.
- 868 **Wosniack ME**, Kirchner JH, Chao LY, Zabouri N, Lohmann C, Gjorgjieva J. Adaptation of spontaneous activ-
869 ity in the developing visual cortex. *eLife*. 2021 Mar; 10:e61619. <https://doi.org/10.7554/eLife.61619>, doi:
870 [10.7554/eLife.61619](https://doi.org/10.7554/eLife.61619), publisher: eLife Sciences Publications, Ltd.
- 871 **Yamins DLK**, Hong H, Cadieu CF, Solomon EA, Seibert D, DiCarlo JJ. Performance-optimized hierarchical models
872 predict neural responses in higher visual cortex. *Proceedings of the National Academy of Sciences*. 2014
873 Jun; 111(23):8619–8624. <http://www.pnas.org/doi/10.1073/pnas.1403112111>, doi: [10.1073/pnas.1403112111](http://www.pnas.org/doi/10.1073/pnas.1403112111),
874 publisher: Proceedings of the National Academy of Sciences.

Supplemental Material

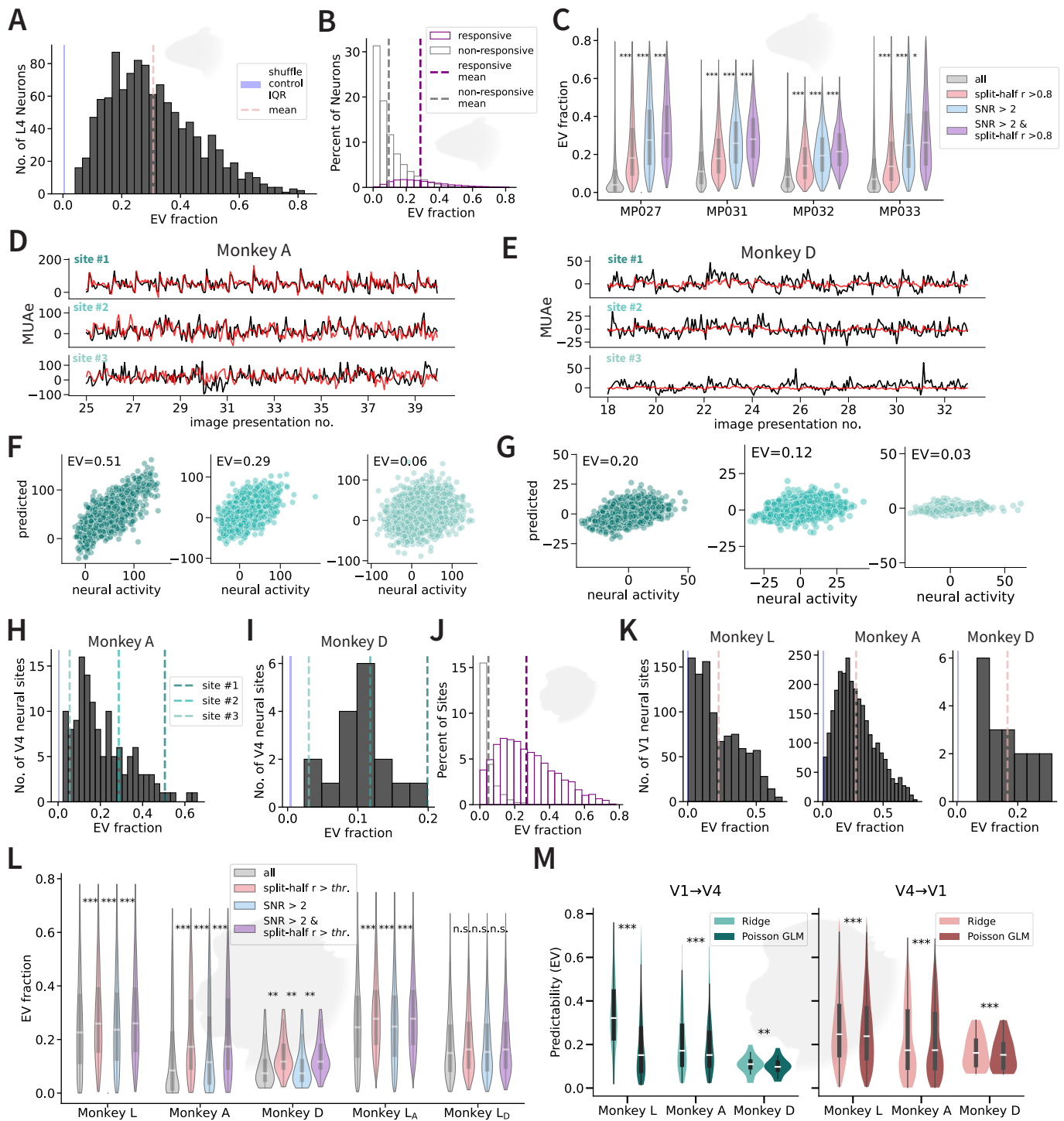


Figure Supplement 1. EV fraction in mouse L4 neurons and monkey V1 neuronal sites and comparison between visual and non-visual neurons/sites. **A.** Distribution of EV fraction in L2/3→L4 regressions in cells deemed visually reliable in 4 mice and 7 recording days ($n = 1113$ neurons). **B.** Distribution of EV fraction values for visually (purple) and non-visually (gray) reliable neurons in mouse L2/3 and L4. In mice, we used a conservative criterion to select neurons that were visually reliable, based on an average signal-to-noise ratio over 2 and a split-half correlation value of at least 0.8 (Methods). **C.** Differences in EV fraction using

different filtering methods to determine visually reliable neurons in mouse L2/3 and L4 across the 4 mice. **D.** Example of monkey A MUAe activity (black) in response to a full-field checkerboard image in three V4 neuronal sites along with regression-model predictions (red) for a typical site (2, middle), site in the top 10% percentile of predictability (1, top), and bottom 10% percentile (3, bottom). **E.** Same as **D** for monkey D. **F.** Predicted neuronal activity versus actual neuronal activity in response to checkerboard and moving bar presentations for monkey A sites 1, 2, and 3 shown in **D**. Each point represents activity in one 2x5 ms timepoint during the 400 ms presentation. **G.** Same as **F** for monkey D neuronal sites shown in **E**. **H.** Distribution of EV fraction in V1→V4 regressions of neural activity in response to visual stimuli in sites deemed visually reliable in monkey A (3 recording days, 30–44 V4 sites recorded per day; $n = 132$ total site recordings). **I.** Same as **H** for monkey D (2 recording days, 7–10 V4 sites recorded per day; $n = 17$ total site recordings). **J.** Distribution of EV fraction for visually (purple) and non-visually (gray) reliable neurons in monkey V1 and V4. Both SNR over 2 and a split-half correlation value of over 0.8 were used to define a neuron to be visually reliable in monkeys L and A. In monkey D, a lower split-half correlation value of 0.6 was used to increase the site count. **K.** Distribution of EV fraction in V4→V1 regressions of neural activity in response to visual stimuli in sites deemed visually reliable in monkeys L (5 recording days, 537–592 V1 sites recorded per day; $n = 2789$ total site recordings), A (3 recording days, 251–381 V1 sites recorded per day; $n = 989$ total site recordings), and D (2 recording days, 8–10 V1 sites recorded per day; $n = 18$ total site recordings). **L.** Differences in EV fraction using different filtering methods to determine visually reliable neurons in macaque V4 and V1 across three monkeys (L, A, and D) V1 and V4 sites and two subsampled permutations of monkey L (to compare to site counts in monkey A and D). Split-half correlation value *thr* represents either 0.8 (monkeys L, A, and L_A) or 0.6 (monkey D and L_D). **M.** Head-to-head comparison of ridge regression EV and Poisson GLM EV on monkey MUAe for V1→V4 (left) and V4→V1 (right). Models share identical train/test folds, 25 ms bins, and temporal gaps; the Poisson GLM enforces non-negativity via a log link on raw MUAe (no baseline subtraction).

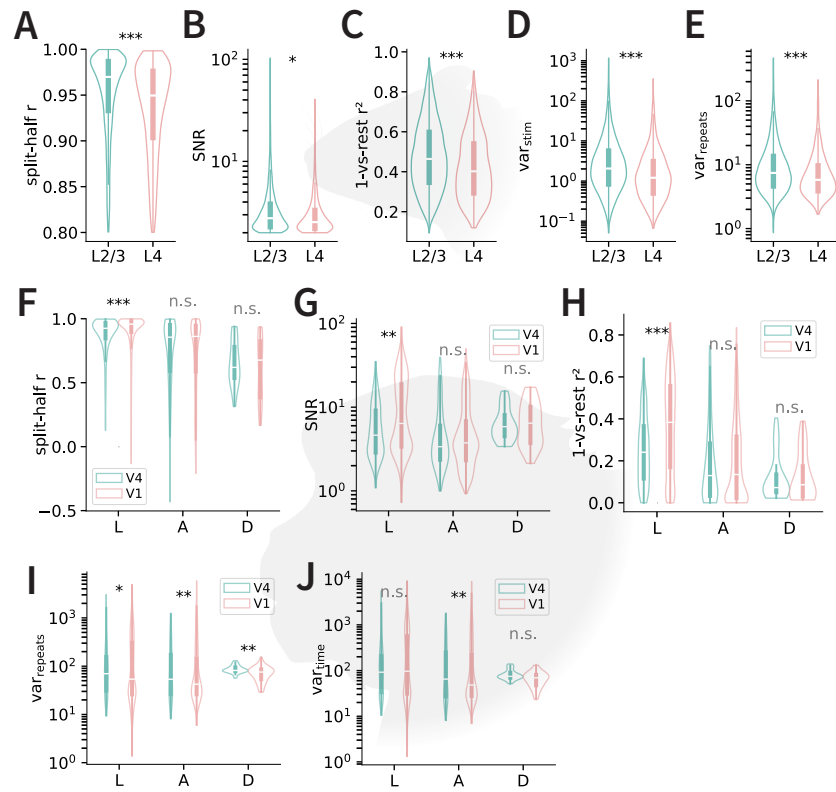


Figure Supplement 2. Neuronal property differences between areas in mouse and monkeys. **A-E.** Differences in self-consistency, SNR, 1-vs-rest squared, variance across stimuli, and variance across repeats between entire visually reliable neuronal populations in mouse L2/3 and L4. **F-J** Differences in self-consistency, SNR, 1-vs-rest squared, variance across time (within repeat), and variance across repeats (within timepoint) between entire visually reliable neuronal populations in monkey V1 and V4.

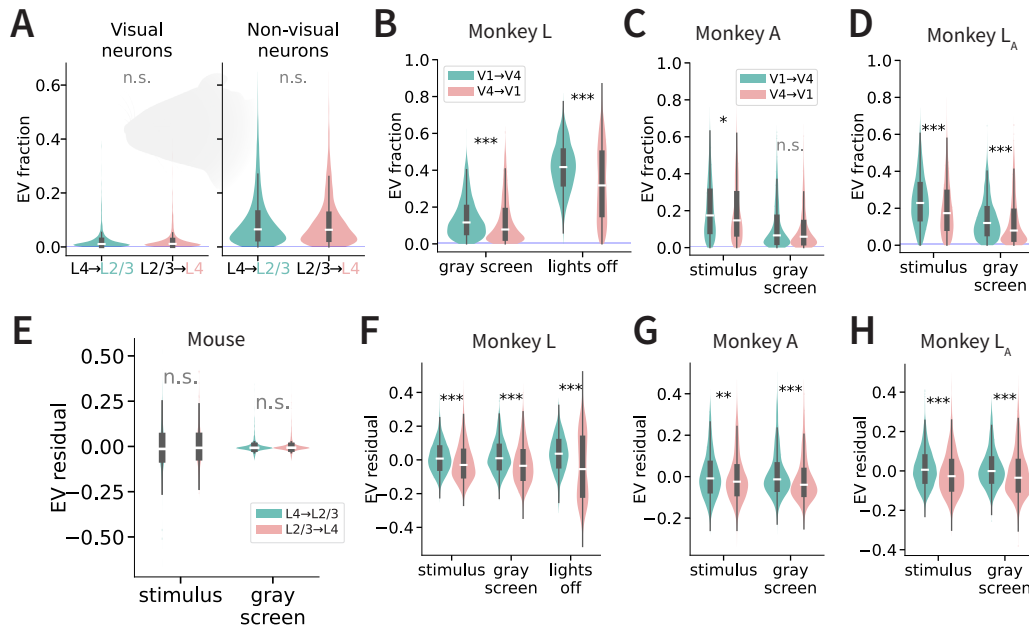


Figure Supplement 3. Differences in monkey inter-cortical predictability directions and lack of difference in mouse inter-laminar predictability directions are also seen in the absence of a stimulus. **A.** Differences in inter-laminar predictability directions in mouse neuronal activity during gray screen presentation in visual (left) and non-visual (right) neurons. **B.** Differences in inter-cortical predictability directions in monkey L during gray screen presentation and during lights off conditions. **C.** Differences in inter-cortical predictability directions in monkey A during stimulus and gray screen presentation. **D.** Same as **C** but for monkey L, after subsampling to match the number of sites in monkey A (L_A). **E.** Differences in EV residuals after removing target-population covariates in mouse neural activity during stimulus and gray screen presentation (self-consistency, SNR, one-vs-rest r^2 , and variance metrics; for details see [Methods](#)). **F** Same as **E** but for monkey L EV residuals during stimulus, gray screen, and lights off conditions. **G.** Same as **E** but for monkey A EV residuals during stimulus and gray screen conditions. **H.** Same as **G** but for monkey L, after subsampling to match the site counts of monkey A (L_A).

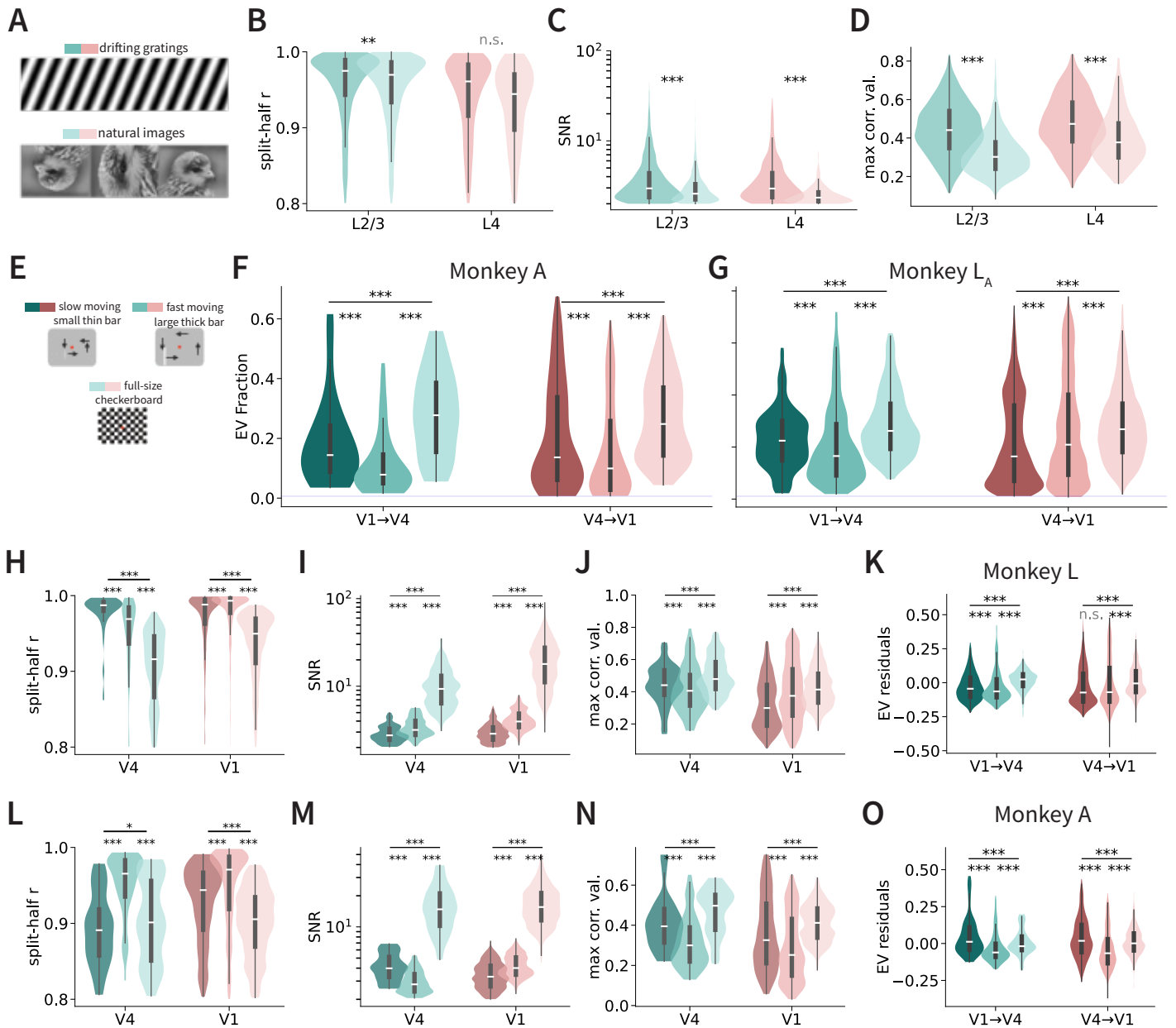


Figure Supplement 5. Neuronal activity properties for different stimulus types. **A.** Sample stimuli for mouse drifting grating and static natural images. **B-D.** Split-half correlation (**B**), SNR (**C**), and maximum correlation values (**D**) for each mouse layer and stimulus type (see color scheme in **A**). **E.** Sample stimuli for monkeys: full-size checkerboard, slow and fast moving bars. **F.** Across-area predictability for each stimulus type (dark: slow bars, medium: fast bars, light: checkerboard) and direction for monkey A. **G.** Same as **F** but for subsampled monkey L (L_A). **H-I.** Same as **B-D** for monkey L V1 and V4 (see color map for each stimulus condition in **E**). **K.** EV residuals for monkey L after regressing, within direction, on SNR, split-half reliability, variance across time (within stimulus), and variance across trials (within timepoint). **L-N.** Same as **B-D** but for monkey A V1 and V4. **O.** Same as **K** but for monkey A.

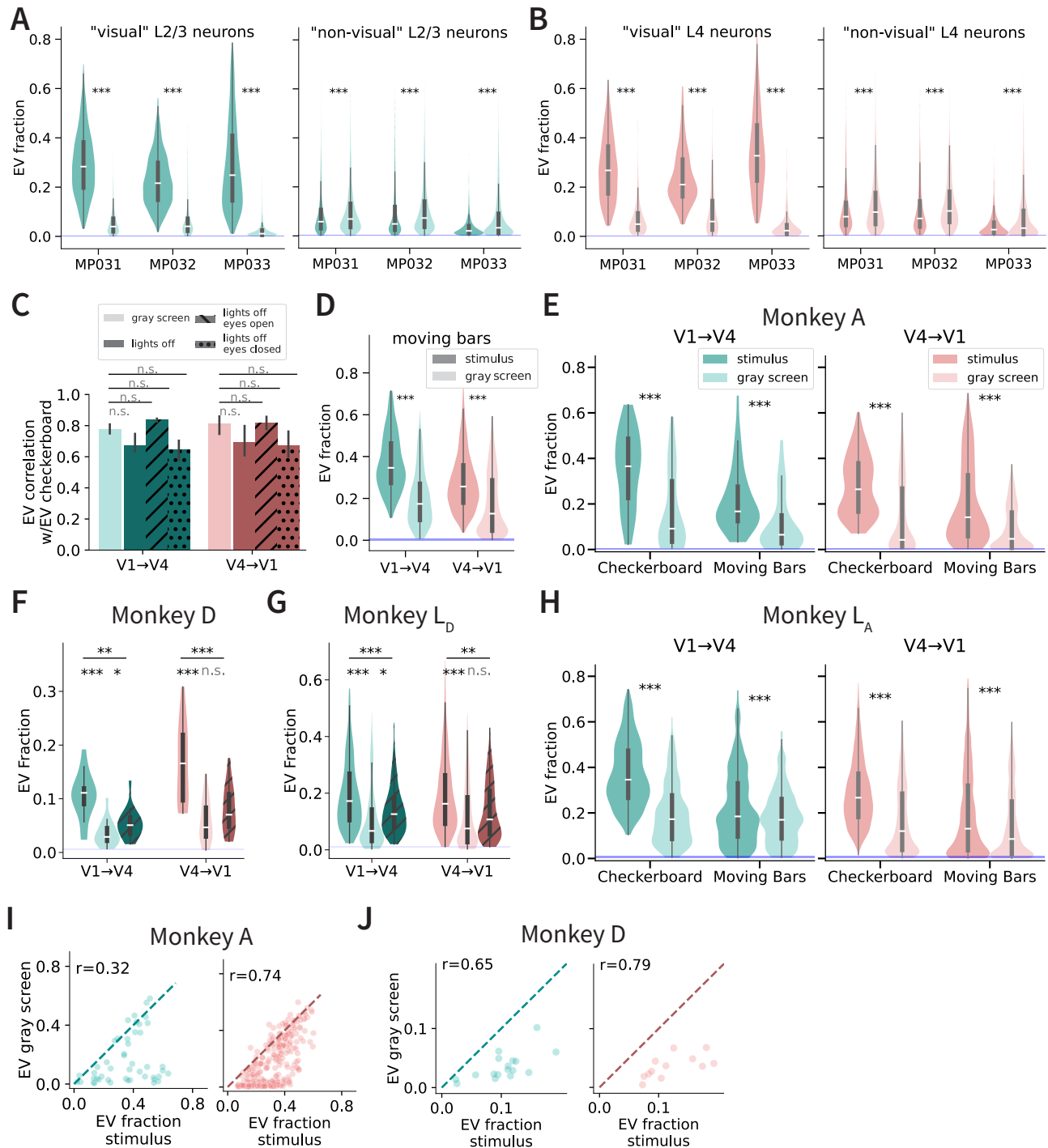


Figure Supplement 6. Comparing stimulus presentation vs. gray screen activity predictions in mouse and monkey. **A.** Differences in inter-laminar predictability between stimulus presentation and gray screen presentation neuronal activity in L4→L2/3 predictions across the three different mice (MP027 did not undergo gray screen presentation recordings). Left: visually reliable L2/3 neurons. Right: non-visually reliable L2/3 neurons. **B.** Same as **A**, but for mouse L2/3→L4 predictions. **C.** Correlation coefficient values between checkerboard presentation and gray screen and lights off conditions in monkey inter-cortical predictability. **D.** Differences in inter-cortical predictability between moving bar presentation and gray screen activity in monkey L V1→V4 and V4→V1 predictions (paired permutation test). **E.** Differences in inter-cortical predictability between stimulus and gray screen presentation across both checkerboard and moving bar stimuli in monkey A. **F.** EV differences in inter-cortical predictability between stimulus, gray screen, and lights off conditions in monkey D. **G.** Same as **F** but for monkey L subsampling sites to match the num-

ber of sites in monkey D (L_D). **H.** Same as **E** but for monkey A, after subsampling sites to match the number of sites in monkey A (L_A) **I.** Correlation between EV in responses to gray screen (y-axis) versus stimulus presentation (x-axis) in monkey A visually reliable neurons (V1→V4:left, green; V4→V1:right, coral). The diagonal line represents the line of equality ($y=x$). r is the Pearson's correlation coefficient. **J.** Same as **I** for monkey D.

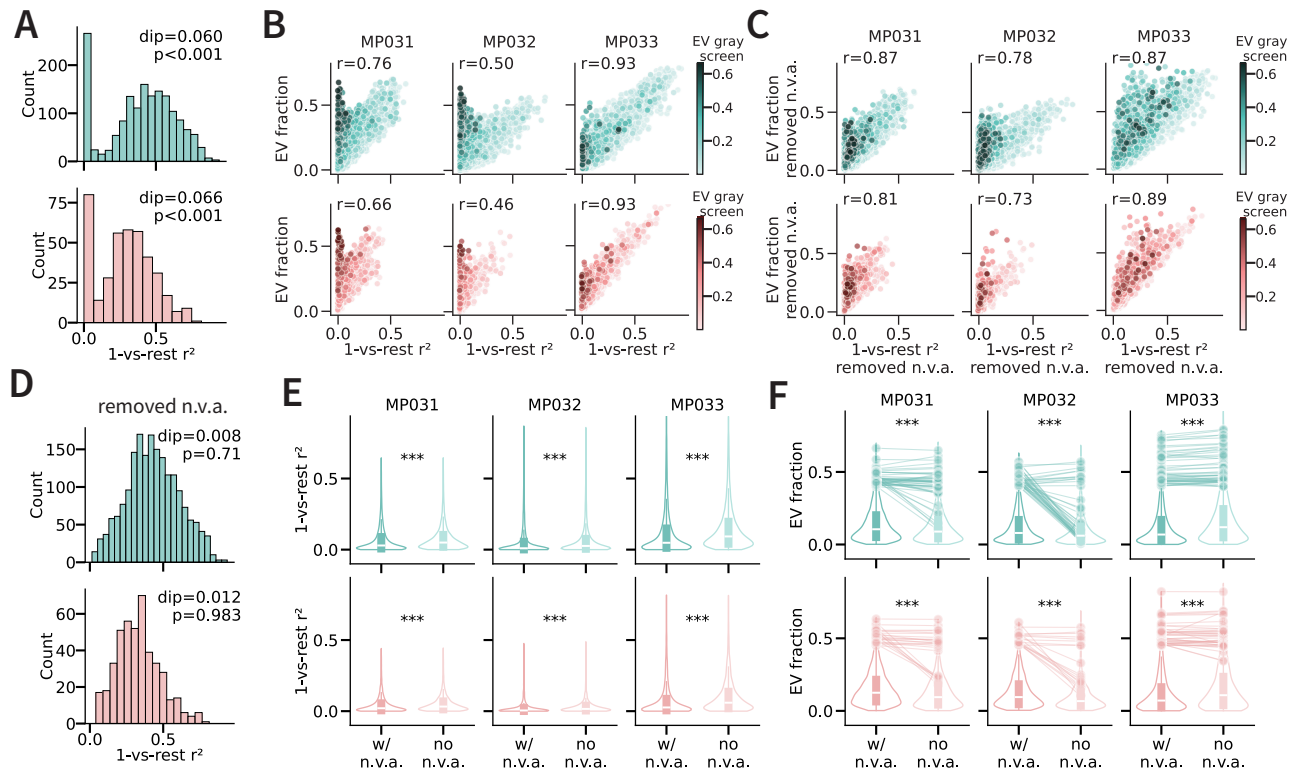


Figure Supplement 7. Properties of visual and nonvisually active neurons in mice. **A.** Bimodal distribution of 1-vs-rest squared correlation values in highly predictable L2/3 (top) and L4 (bottom) neurons ($EV > 0.4$). Hartigan's dip test was applied to test bimodality (top right corner). **B.** Relationship between 1-vs-rest squared correlation and EV fraction in L2/3 (top, green) and L4 (bottom, coral) neurons in three mice (columns). **C.** Same as **B** after projecting out "non-visual ongoing" activity (*Stringer et al., 2019a*), see text for details. **D.** Distribution of 1-vs-rest squared correlation values in highly predictable L2/3 (top) and L4 (bottom) neurons ($EV > 0.4$) after projecting out "non-visual ongoing" activity. **E.** One-vs-rest squared correlation values when including (left) and not including (right) non-visual ongoing activity in each mouse (columns). * denote paired permutation test. **F.** EV fraction when including (left) and not including (right) non-visual ongoing activity dimensions in L2/3 (top, green) and L4 (bottom, coral) in each mouse (columns). Sample subset of neurons with initial prediction values of over 0.4 visualized with lineplots. * denote paired permutation test.

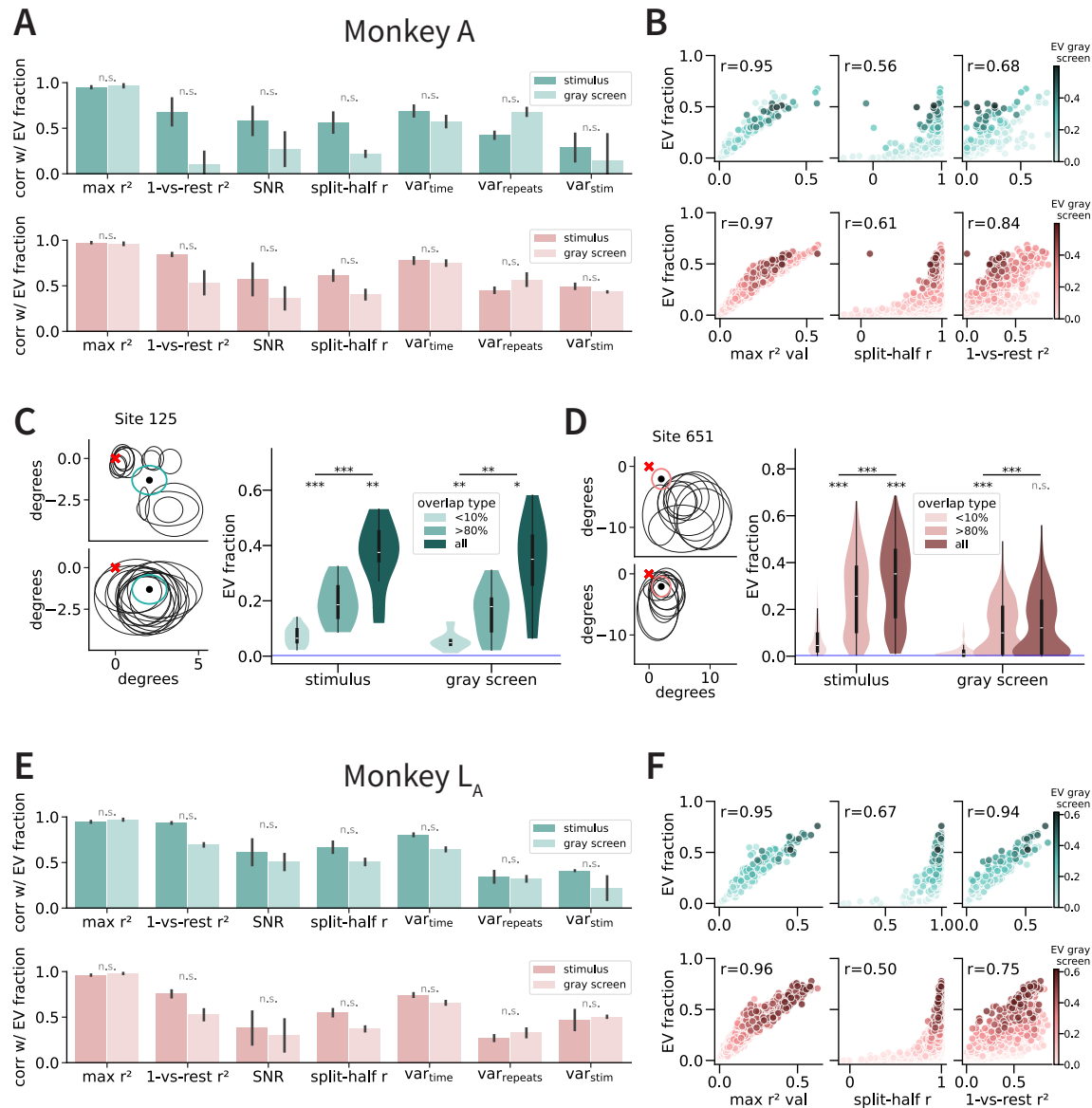


Figure Supplement 8. Influence of SNR, variance, and receptive field overlap is consistent across monkeys L and A. **A.** Correlation between different properties with the predictability of V4 (green) and V1 (coral) neuronal responses during the presence (dark color) or absence (light color) of visual stimulus in monkey A. **B.** Relationship between the EV fraction and three neuronal properties in sites for monkey A V4 (green) and V1 (coral) responses to visual stimuli. Hue represents the degree of predictability for the same sites during gray screen presentations (see color scale on right). **C.** Left, top: Receptive fields of one sample V4 neuronal site in monkey A (green circle, array 2 electrode 187) and 14 randomly selected V1 neuronal sites as predictors (black circles), constrained on sites that share less than 10% receptive field overlap with the V4 site. Bottom: Receptive fields of the same neuronal site 125 and 14 randomly selected V1 neuronal sites used as predictors, constrained on sites that share at least 80% receptive field overlap with the V4 site. Right: EV fraction of V4 neural activity ($n = 18$ site recordings per activity type) using 14 V1 predictor sites with less than 10% RF overlap (light green), 14 predictor sites with at least 80% RF overlap (green), and all predictors (dark green). Predictions were computed for recordings in response to the stimulus presentation (sliding bars and full-field checkerboard images) and gray screen presentation. **D.** Bottom and top left: Same as **C** but for monkey A sample V1 site 651. Right: Same as **C**, but for V1 ($n = 378$ site recordings per activity type). Instead of 14, 10 prediction sites were used to predict V1 due to low sample count of V4 that fulfilled both types of RF overlap percentages. **E-F.** Same as **A-B** but for monkey L after

subsampling sites to match the number of sites in monkey A (L_A).

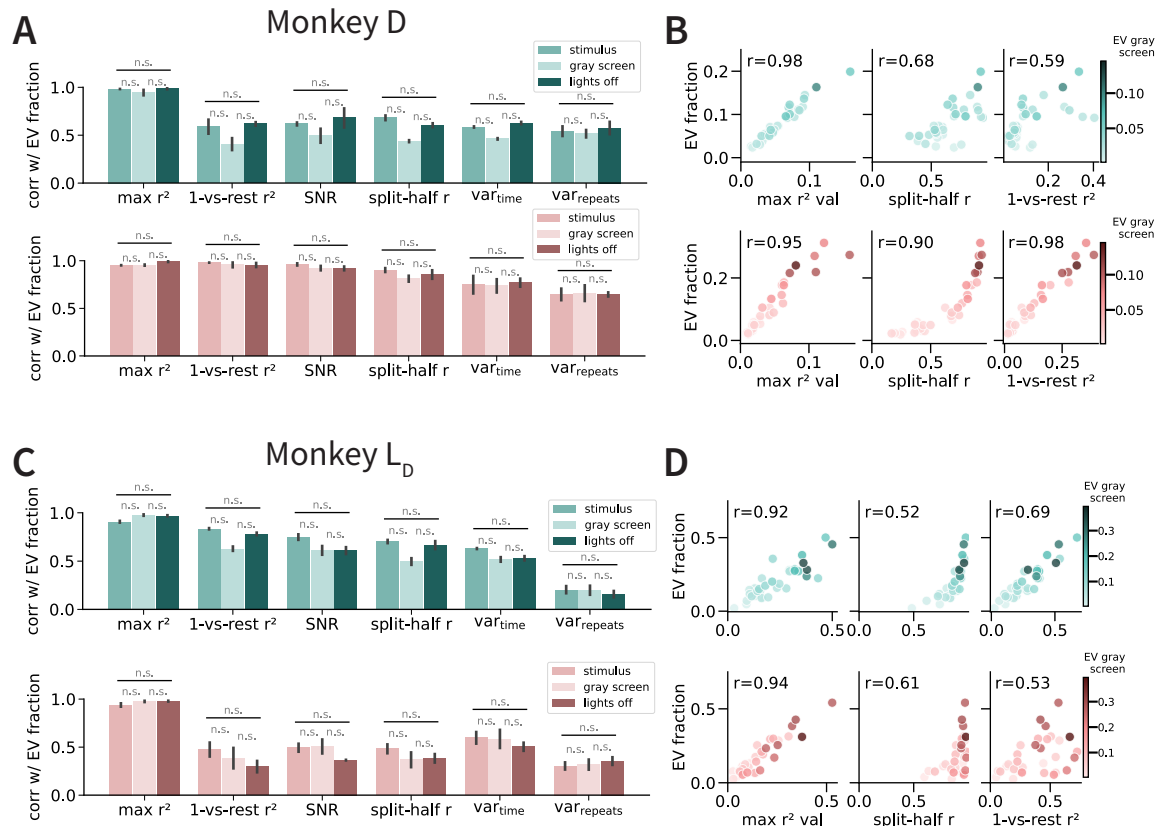


Figure Supplement 9. Influence of SNR, variance, and split-half reliability is consistent across monkeys L and D. **A.** Correlation between different properties with the predictability of V4 (green) and V1 (coral) neuronal responses during the presence (dark color) or absence (light color) of visual stimulus in monkey D. **B.** Relationship between three properties and their predictability in sites for monkey D V4 (green) and V1 (coral) responses from stimuli presentations. Hue represents the degree of predictability for the same sites during gray screen presentations. **C–D.** Same as A–B but for subsample monkey L_D .

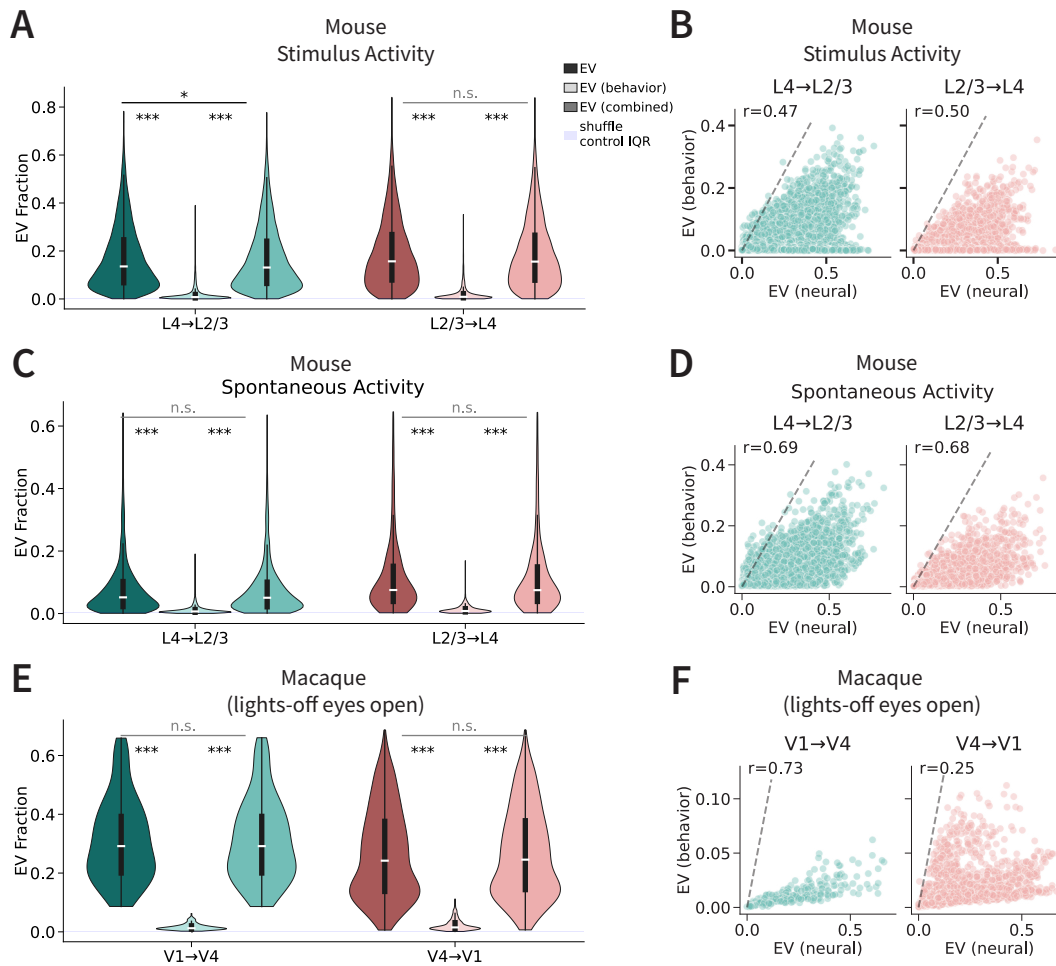


Figure Supplement 10. Behavioral contributions to inter-area predictability in mouse and monkey. **A.** Distribution of EV fraction for neural-only (dark colors), behavior-only (face-motion SVD and running speed; light colors), and combined models (medium colors) in mouse L4→L2/3 and L2/3→L4 predictions during stimulus activity. **B.** Scatter plots comparing EV from behavior-only models (y-axis) versus neural-only models (x-axis) for mouse L2/3 response prediction (left) and mouse L4 response prediction (right). **C–D.** Same as **A–B**, but for spontaneous activity in mouse. **E.** Distribution of EV fraction for neural-only, pupil-only, and combined models in monkey L V1→V4 and V4→V1 predictions during resting state with eyes open. **F.** Scatter plots comparing EV from behavior-only models (y-axis) versus neural-only models (x-axis) for monkey V4 response prediction (left) and monkey V1 response prediction (right). The dashed line represents the line of equality ($y=x$).

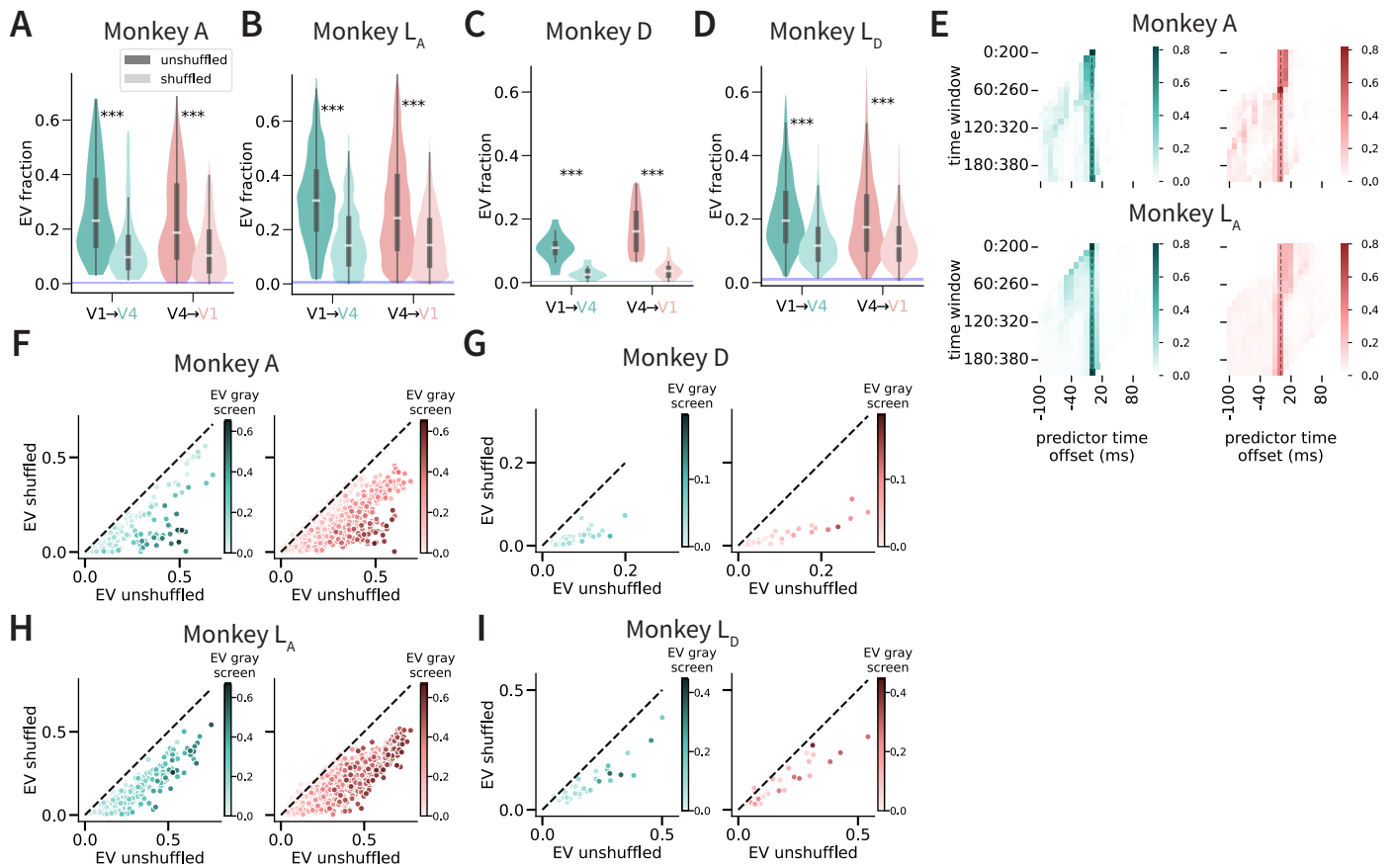


Figure Supplement 11. Time-dependent effects on EV predictability in monkeys A and D. **A-D** Distribution of EV fraction in unshuffled (dark) and shuffled (light) trial-repeat activity in monkey A, monkey L subsampled to match the number of sites in monkey A (L_A), monkey D, and monkey L subsampled to match the number of sites in monkey D (L_D). * denote paired permutation tests. **E.** Time offset prediction results across both V1→V4 (left, green) and V4→V1 (right, coral) prediction directions in monkeys A (top) and subsampled L_A (bottom). Each square corresponds to the fraction of neuronal sites whose neural activity was best predicted during that offset period and time window. **F-I.** Relationship between shuffled (y-axis) and unshuffled (x-axis) trial repeat EVs in V1 → V4 (left, green) and V4 → V1 (right, coral) directions in monkeys A, subsampled L_A , D, and subsampled L_D . Hue represents EV fraction during gray screen activity (see color scale on right).

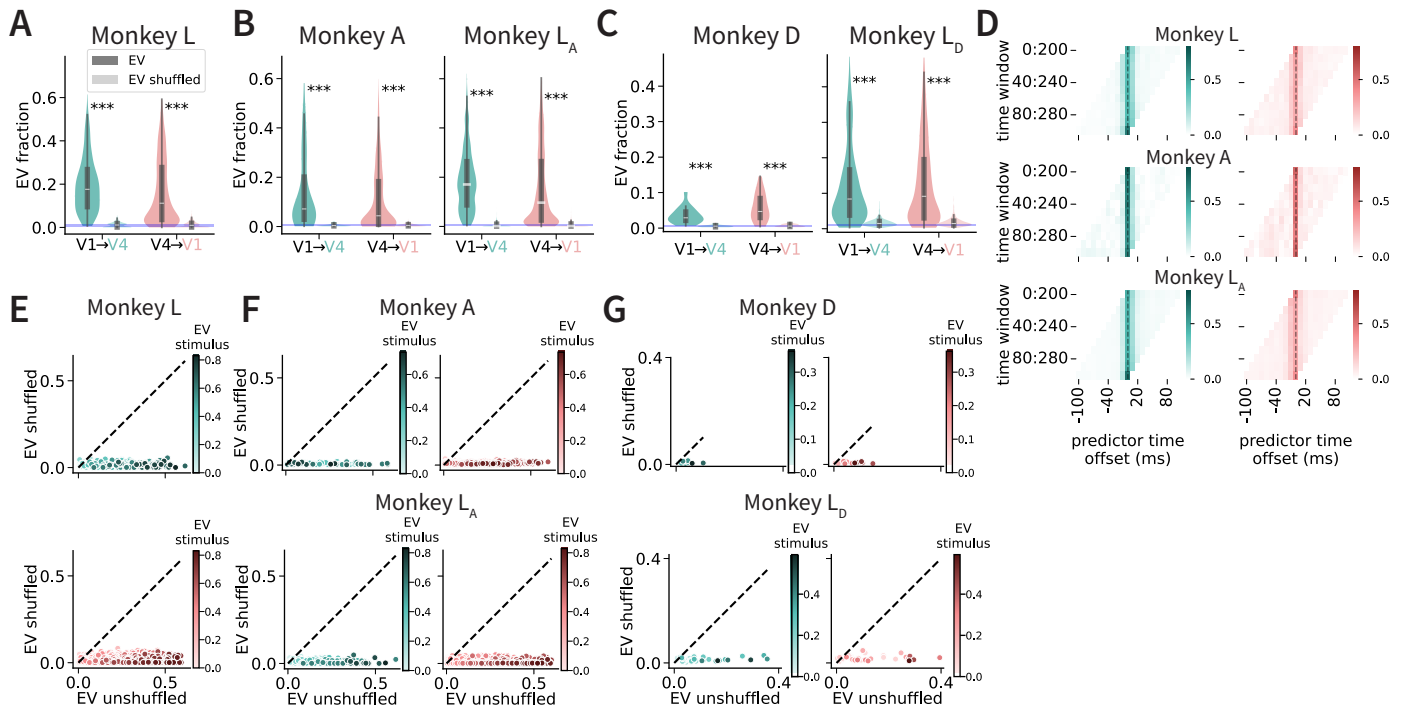


Figure Supplement 12. Time-dependent effects on EV predictability during spontaneous conditions. Distribution of EV fraction for unshuffled (dark) and shuffled (light) trial-repeat activity during gray screen presentations (paired permutation test) in monkeys L, A, subsampled L_A, D, and subsampled L_D. * denote paired permutation tests. **D**. Time offset prediction results across both V1→V4 (left, green) and V4→V1 (right, coral) prediction directions during gray screen presentations in monkeys A (top) and subsampled L_A (bottom). Each square corresponds to the fraction of neuronal sites whose neural activity was best predicted during that offset period and time window. **E-G**. Relationship between shuffled (y-axis) and unshuffled (x-axis) trial repeat EVs in V1 → V4 (left, green) and V4 → V1 (right, coral) directions in monkeys L (**E**), A & subsampled L_A (**F**), and D & subsampled L_D (**G**). Hue represents EV fraction during stimulus activity (see color scale on right).

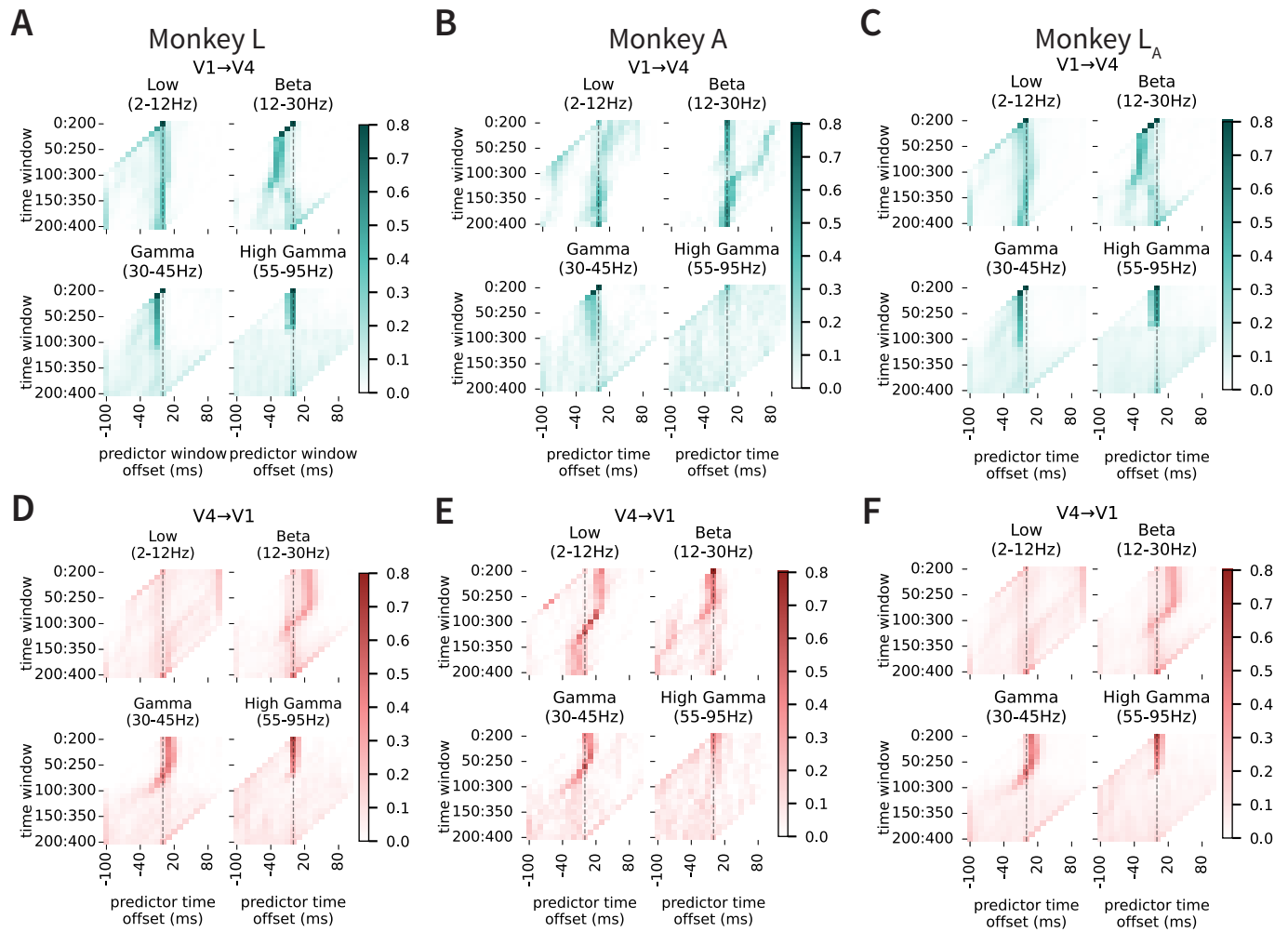


Figure Supplement 13. Time-dependent effects on EV predictability across LFP bands in monkey V1/V4. A-C. Time offset prediction results across both V1→V4 (left, green) and V4→V1 (right, coral) prediction directions in monkeys L, A, and L_A. Columns correspond to band-limited LFP amplitude (Hilbert envelope) from Low (2–12 Hz), Beta (12–30 Hz), Gamma (30–45 Hz), and High-gamma (55–95 Hz). LFP preprocessing included removal of narrow line artifacts (notch filter at 50 Hz and harmonics), band-pass filtering, and Hilbert amplitude extraction; envelopes were z-scored per unit. The dashed vertical line marks zero offset. **D-F.** Same as **A–B**, but predicting V1 from V4.